

EMERGING FUNGAL PATHOGENS: CHALLENGES, MECHANISMS, AND NOVEL ANTIFUNGAL STRATEGIES

Md. Al Amin^{1,3*}, Moazzema Binta Bashar², Sree Karma Tigga³, Arin Akther Purne¹, Md. Rezwan Hossain³

¹Department of Pharmacy, University of Information Technology and Sciences (UITS), Dhaka 1212, Bangladesh.

²State University of Bangladesh, Dhaka-1461, Bangladesh.

³Department of Pharmacy, University of Rajshahi, Rajshahi-6205, Bangladesh.

Article Received: 01 March 2026

Article Review: 23 March 2026

Article Accepted: 14 April 2026

*Corresponding Author: Md. Al Amin

Department of Pharmacy, University of Information Technology and Sciences (UITS), Dhaka 1212, Bangladesh.

DOI: <https://doi.org/10.5281/zenodo.19923319>

How to cite this Article: Md. Al Amin, Moazzema Binta Bashar, Sree Karma Tigga, Arin Akther Purne, Md. Rezwan Hossain (2026). EMERGING FUNGAL PATHOGENS: CHALLENGES, MECHANISMS, AND NOVEL ANTIFUNGAL STRATEGIES. World Journal of Pharmacy and Medical Science, 2(5): 26-47.



Copyright © 2026 Md. Al Amin | World Journal of Pharmacy and Medical Science

This is an open-access article distributed under creative Commons Attribution-NonCommercial 4.0 International license ([CC BY-NC 4.0](https://creativecommons.org/licenses/by-nc/4.0/))

ABSTRACT

Emerging fungal pathogens represent a growing global health threat, particularly affecting immunocompromised populations including individuals with HIV/AIDS, malignancies, organ transplants, and those receiving immunosuppressive therapies. The incidence of invasive and opportunistic mycoses has risen markedly due to clinical and environmental factors. Pathogens such as *Candida auris*, azole-resistant *Aspergillus fumigatus*, *Cryptococcus* spp., and Mucorales demonstrate multidrug resistance, high virulence, and the ability to cause severe, often fatal infections. This review was conducted through a structured literature survey of peer-reviewed articles, reviews, and authoritative sources retrieved from PubMed, Scopus, Web of Science, and Google Scholar over a decade from 2014 to 2026. Search terms included "emerging fungal pathogens," "antifungal resistance," "novel antifungals," and "fungal diagnostics." Data were extracted on pathogen identification, resistance mechanisms, host-pathogen interactions, virulence factors, novel therapeutic strategies, and diagnostic advances. Resistance mechanisms identified include drug target alterations, efflux pump overexpression, biofilm formation, stress response activation, genetic adaptability, metabolic rewiring, and cell wall remodeling—often acting synergistically to reduce antifungal efficacy. Virulence is enhanced through adhesion, tissue invasion, enzymatic degradation, immune evasion, and morphological plasticity. Current antifungal therapies remain limited to azoles, echinocandins, and polyenes, each with notable toxicity, resistance issues, and diagnostic challenges. To address these limitations, novel strategies under development include new antifungal agents targeting unique pathways (ibrexafungerp, fosmanogepix, olorofim), nanotechnology-based drug delivery, antifungal peptides, immunotherapy, combination therapies, natural product derivatives, and gene- or RNA-based interventions. Diagnostic advances including PCR, MALDI-TOF mass spectrometry, and biomarker detection enable earlier and more accurate identification. This review synthesizes current knowledge on emerging fungal pathogens, their resistance and virulence mechanisms, and innovative antifungal strategies, highlighting critical research gaps and emphasizing the urgent need for integrated surveillance, rapid diagnostics, and effective therapeutic interventions.

KEYWORDS: Emerging fungal pathogens, Antifungal resistance, Multidrug resistance, Virulence mechanisms, Novel antifungal strategies, Pathogen adaptation.

INTRODUCTION

Fungal infections have emerged as a significant and often underrecognized threat to global public health, contributing substantially to morbidity and mortality, particularly among immunocompromised individuals. In recent decades, the incidence of invasive and

opportunistic mycoses has increased markedly, driven by the expanding population of patients with weakened immune systems, including those with HIV/AIDS, malignancies, organ transplants, and individuals receiving immunosuppressive or broad-spectrum antimicrobial therapies.^[1] Despite their clinical

importance, fungal diseases have historically received less attention compared to bacterial and viral infections, resulting in limited therapeutic options and slower progress in antifungal drug development.^[2]

The term “emerging fungal pathogens” refers to previously unrecognized, newly evolved, or rapidly spreading fungal species that pose a growing risk to human health. Among these, multidrug-resistant organisms such as *Candida auris* have gained particular attention due to their ability to cause nosocomial outbreaks, persist in healthcare environments, and exhibit resistance to multiple antifungal drug classes.^[3] Similarly, *Aspergillus fumigatus* has demonstrated increasing resistance to azole antifungals, often linked to environmental exposure to agricultural fungicides.^[4] Other clinically important fungi, including *Cryptococcus neoformans*, *Mucorales*, *Fusarium*, and *Scedosporium* species, are also associated with severe and often life-threatening infections, especially in vulnerable populations.^[5]

Several factors contribute to the emergence and spread of these fungal pathogens. Climate change has been proposed as a key driver, enabling environmental fungi to adapt to higher temperatures and potentially overcome the thermal barrier of the human host.^[6] Additionally, widespread antifungal use in both clinical and agricultural settings has accelerated the selection of resistant strains.^[7] The intrinsic complexity of fungal cells, which share many similarities with human eukaryotic cells, further complicates antifungal drug development by limiting the availability of selective therapeutic targets.^[7]

Managing fungal infections presents multiple challenges. The current antifungal armamentarium is restricted to a few major drug classes—azoles, echinocandins, and polyenes—each associated with limitations such as toxicity, drug–drug interactions, and emerging resistance.^[8] Diagnostic delays due to nonspecific clinical manifestations and limitations of conventional laboratory methods further exacerbate disease outcomes.^[9] Moreover, the ability of many fungi to form biofilms and undergo morphological transitions enhances their survival, virulence, and resistance to treatment.^[10]

Given these challenges, there is an urgent need to better understand the mechanisms underlying fungal pathogenicity and antifungal resistance, as well as to develop innovative therapeutic strategies. Recent advances in molecular biology, immunology, and nanotechnology have opened new avenues for antifungal intervention, including the development of novel drug candidates, immunotherapeutic approaches, and combination therapies aimed at improving treatment efficacy and overcoming resistance.

This review aims to provide a comprehensive overview of emerging fungal pathogens, focusing on their

epidemiology, mechanisms of resistance and virulence, and the current challenges in clinical management. Furthermore, it highlights recent progress in the development of novel antifungal strategies and identifies key research gaps that must be addressed to effectively combat the growing burden of fungal infections.

METHODOLOGY

This review was conducted through a structured literature survey to analyze emerging fungal pathogens, their resistance mechanisms, virulence factors, and novel therapeutic strategies. Peer-reviewed articles, reviews, and authoritative sources from 2014 to 2026 were retrieved from databases including PubMed, Scopus, Web of Science, and Google Scholar using search terms such as “emerging fungal pathogens,” “antifungal resistance,” “*Candida auris*,” “*Aspergillus* resistance,” “novel antifungals,” “antifungal peptides,” “nanotechnology antifungal,” and “fungal diagnostics.” Studies were included if they focused on clinical or environmental fungal pathogens, resistance mechanisms, emerging trends, novel therapeutic approaches (drugs, immunotherapy, natural products, or nanotechnology), or diagnostic advances, while non-English publications, inaccessible full texts, or irrelevant reports were excluded. Data were extracted on pathogen identification and epidemiology, molecular mechanisms of resistance, host–pathogen interactions, virulence factors, novel antifungal strategies, and diagnostic developments, including surveillance and One Health considerations. The information was synthesized narratively, integrating experimental evidence with clinical observations to provide a comprehensive overview of current challenges, mechanistic insights, and future directions in the management of emerging fungal infections.

1. Major Emerging Fungal Pathogens

The spectrum of clinically significant fungal pathogens has expanded considerably in recent years, with several species demonstrating increased virulence, geographic spread, and antifungal resistance. These emerging fungi are responsible for a wide range of infections, from superficial diseases to life-threatening systemic mycoses, particularly in immunocompromised hosts. **Table 1** represents an overview of emerging fungal pathogens and their clinical significance.

1.1 *Candida* Species

Species of the genus *Candida* remain the leading cause of invasive fungal infections worldwide. While *Candida albicans* has historically been the predominant pathogen, there has been a notable epidemiological shift toward non-albicans *Candida* species with distinct resistance profiles.^[11]

Candida auris has emerged as a major global health concern due to its multidrug resistance, high transmissibility, and ability to persist on hospital surfaces, leading to outbreaks in healthcare settings.^[3] It is frequently resistant to azoles, polyenes, and

occasionally echinocandins, severely limiting treatment options.^[12] In addition to *C. auris*, other non-albicans species such as *C. glabrata*, *C. tropicalis*, *C. parapsilosis*, *C. krusei*, and *C. lusitaniae* are increasingly reported. These species often exhibit reduced susceptibility to commonly used antifungal agents and are associated with bloodstream infections, biofilm formation on medical devices, and increased mortality rates.^[13,14]

1.2 *Aspergillus* Species

The genus *Aspergillus* comprises several opportunistic pathogens, with *Aspergillus fumigatus* being the most clinically significant species responsible for invasive aspergillosis.^[15] This pathogen primarily affects immunocompromised individuals, including patients with hematological malignancies and those undergoing organ transplantation.^[16]

A major concern is the rising incidence of azole-resistant *A. fumigatus*, which compromises first-line antifungal therapy. Resistance is often linked to mutations in the *cyp51A* gene, and notably, environmental exposure to azole fungicides used in agriculture has been identified as a key driver of resistance development.^[17,18] Other clinically relevant species include *A. flavus*, *A. niger*, *A. terreus*, and *A. nidulans*, each associated with distinct clinical manifestations and varying antifungal susceptibility patterns.^[19,20]

1.3 *Cryptococcus* Species

Cryptococcus neoformans and *Cryptococcus gattii* are encapsulated yeasts that cause severe systemic infections, particularly cryptococcal meningitis.^[21] These infections are a leading cause of mortality among individuals with advanced HIV/AIDS and other immunocompromising conditions.^[22]

Cryptococcus neoformans is globally distributed and predominantly affects immunocompromised patients, whereas *C. gattii* can infect immunocompetent individuals and is associated with outbreaks in specific geographic regions.^[23] The ability of these pathogens to cross the blood–brain barrier and establish infection in the central nervous system (CNS) is a key virulence feature.^[24] Challenges in management include limited therapeutic options, prolonged treatment duration, and high relapse rates.

1.4 Other Emerging Fungi

Beyond the major genera previously discussed, several additional fungal groups are increasingly recognized as opportunistic pathogens, often associated with high mortality and intrinsic resistance to antifungal therapy.

Mucorales (Order Mucorales)

This group includes *Rhizopus*, *Mucor*, *Lichtheimia*, *Rhizomucor*, and *Cunninghamella* species, which are responsible for mucormycosis—a rapidly progressing

and frequently fatal infection.^[25] Cases of mucormycosis have surged in patients with COVID-19, particularly those with diabetes or corticosteroid treatment.^[26]

Hyaline hyphomycetes

Species such as *Fusarium*, *Scedosporium*, *Lomentospora prolificans*, *Paecilomyces*, and *Acremonium* are emerging as clinically significant pathogens. They can cause disseminated infections, keratitis, and onychomycosis, and often display resistance to multiple antifungal agents.^[27,28]

Dematiaceous (melanized) fungi

This group includes *Cladophialophora*, *Exophiala*, *Alternaria*, *Bipolaris*, and *Curvularia* species, which are responsible for phaeohyphomycosis. The melanin in their cell walls enhances virulence and contributes to resistance against host immune defenses.^[29,30]

Dimorphic fungi

Thermally dimorphic fungi such as *Histoplasma capsulatum*, *Blastomyces dermatitidis*, *Coccidioides immitis*, *Paracoccidioides brasiliensis*, and *Talaromyces marneffeii* are increasingly reported outside their traditional endemic regions, driven by travel and climate change.^[31,32]

Other emerging fungi

Additional opportunistic pathogens include *Trichosporon* spp., *Malassezia* spp. (linked to bloodstream infections in neonates and immunocompromised patients), *Geotrichum* spp., and *Pneumocystis jirovecii*, the latter being a leading cause of pneumonia in immunosuppressed individuals.^[33]

Collectively, the growing diversity of fungal pathogens, coupled with their evolving resistance mechanisms and expanding ecological niches, underscores the urgent need for improved surveillance, rapid diagnostics, and the development of effective antifungal therapies.

Table 1: Emerging fungal pathogens, their notable species, associated diseases, and populations at risk.

Fungal Pathogen	Emerging / Notable Species	Associated Diseases	At-Risk Population / Notes	Ref
<i>Candida</i> spp.	<i>C. auris</i> , <i>C. glabrata</i> , <i>C. krusei</i> , <i>C. parapsilosis</i>	Candidemia, bloodstream infections, invasive candidiasis	Hospitalized, immunocompromised, ICU patients	[34,35]
<i>Aspergillus</i> spp.	<i>A. fumigatus</i> , <i>A. flavus</i> , <i>A. terreus</i> , <i>A. niger</i>	Invasive aspergillosis, chronic pulmonary aspergillosis	Immunocompromised, transplant recipients, chronic lung disease	[19,36]
<i>Cryptococcus</i> spp.	<i>C. neoformans</i> , <i>C. gattii</i>	Cryptococcal meningitis, pulmonary cryptococcosis	HIV/AIDS, immunosuppressed, organ transplant patients	[37,38]
<i>Mucorales</i>	Rhizopus, Mucor, Lichtheimia, Apophysomyces	Mucormycosis (rhinocerebral, pulmonary, cutaneous)	Diabetics, neutropenic, immunosuppressed	[39,40]
<i>Fusarium</i> spp.	<i>F. solani</i> , <i>F. oxysporum</i>	Fusariosis, keratitis, onychomycosis	Immunocompromised, contact lens users	[41,42]
<i>Scedosporium</i> spp.	<i>S. apiospermum</i> , <i>S. prolificans</i> , <i>S. boydii</i>	Invasive infections, pulmonary, CNS	Immunocompromised, cystic fibrosis patients	[43,44]
<i>Exophiala</i> spp.	<i>E. dermatitidis</i> , <i>E. spinifera</i>	Phaeohyphomycosis, systemic infections	Immunocompromised, sometimes healthy individuals	[45,46]
<i>Emergomyces</i> spp.	<i>E. africanus</i> , <i>E. pasteurianus</i>	Emergomycosis (disseminated fungal infection)	HIV/AIDS, immunocompromised populations	[47,48]
<i>Talaromyces</i> spp.	<i>T. marneffeii</i>	Penicilliosis marneffeii, disseminated infection	HIV/AIDS, Southeast Asia endemic regions	[49,50]
<i>Pneumocystis</i> spp.	<i>P. jirovecii</i>	Pneumocystis pneumonia (PCP)	HIV/AIDS, immunosuppressed, organ transplant patients	[51,52]
<i>Trichosporon</i> spp.	<i>T. asahii</i> , <i>T. inkin</i>	Trichosporonosis, bloodstream infections, fungaemia	Immunocompromised, neutropenic patients	[53,54]
<i>Cladosporium</i> spp.	<i>C. cladosporioides</i> , <i>C. herbarum</i>	Allergic fungal sinusitis, cutaneous infections	Immunocompromised, allergy-prone individuals	[55,56]
<i>Paecilomyces</i> spp.	<i>P. lilacinus</i> , <i>P. variotii</i>	Keratitis, onychomycosis, systemic infections	Immunocompromised, ocular trauma	[57,58]
<i>Blastomyces</i> spp.	<i>B. dermatitidis</i> , <i>B. gilchristii</i>	Blastomycosis (pulmonary and systemic)	Immunocompromised, people in endemic areas (North America)	[59,60]
<i>Histoplasma</i> spp.	<i>H. capsulatum</i>	Histoplasmosis, disseminated infection	Immunocompromised, HIV/AIDS, endemic regions (Americas)	[61,62]
<i>Paracoccidioides</i> spp.	<i>P. brasiliensis</i> , <i>P. lutzii</i>	Paracoccidioidomycosis	Immunocompromised, endemic to South America	[63,64]
<i>Sporothrix</i> spp.	<i>S. schenckii</i> , <i>S. brasiliensis</i>	Sporotrichosis (cutaneous, lymphocutaneous)	Traumatic inoculation, gardeners, immunocompromised	[65,66]
<i>Alternaria</i> spp.	<i>A. alternata</i> , <i>A. infectoria</i>	Cutaneous infections, sinusitis, keratitis	Immunocompromised, chronic sinusitis patients	[67,68]

2. Challenges in Managing Fungal Infections

The effective management of fungal infections remains a significant clinical challenge due to a combination of biological, pharmacological, and diagnostic limitations. Despite advances in medical science, the burden of invasive mycoses continues to rise, particularly among immunocompromised populations, highlighting critical gaps in current antifungal strategies.^[69]

One of the primary challenges is the limited availability of antifungal drug classes. Currently, antifungal therapy relies mainly on three major classes: azoles, echinocandins, and polyenes. These drugs target a relatively narrow range of fungal cellular processes, such as ergosterol synthesis or cell wall integrity. The limited diversity of targets not only restricts therapeutic options

but also facilitates the rapid emergence of resistance, especially with prolonged or prophylactic use.^[70,71]

Closely related to this limitation is the issue of drug toxicity and adverse effects. Among the available antifungals, amphotericin B, a polyene antibiotic, is highly effective but associated with significant nephrotoxicity and infusion-related reactions.^[72] Although lipid-based formulations have improved its safety profile, their high-cost limits accessibility in many resource-constrained settings. Azoles, while generally safer, are linked to hepatotoxicity and significant drug–drug interactions, particularly in patients receiving multiple medications.^[73]

Another major obstacle is the lack of rapid and reliable diagnostic methods. Conventional diagnostic techniques, such as fungal culture and microscopy, are time-consuming and often lack sensitivity, leading to delayed diagnosis and treatment initiation.^[74] Advanced diagnostic tools, including molecular assays and antigen detection tests, are not universally available, particularly in low- and middle-income countries.^[75] This diagnostic delay contributes significantly to poor clinical outcomes.

The ability of many fungi to form biofilms further complicates treatment. Biofilms are structured microbial communities that adhere to surfaces such as medical devices (e.g., catheters, implants) and are encased in an extracellular matrix.^[76] Within biofilms, fungal cells exhibit reduced metabolic activity and increased resistance to antifungal agents, making infections difficult to eradicate and often necessitating device removal.^[77]

Fungal infections are also associated with high mortality rates, particularly in cases of invasive disease such as candidemia, invasive aspergillosis, and cryptococcal meningitis.^[78] Mortality is often exacerbated by delayed diagnosis, limited therapeutic options, underlying comorbidities, and the emergence of drug-resistant strains.^[79]

Finally, the absence of effective vaccines against most fungal pathogens represents a major unmet need. Unlike bacterial and viral infections, for which vaccines have significantly reduced disease burden, no widely approved vaccines are currently available for human fungal diseases.^[80] The development of antifungal vaccines is complicated by the eukaryotic nature of fungi, antigenic complexity, and challenges in eliciting durable protective immunity, particularly in immunocompromised individuals.^[81]

So, the management of fungal infections is hindered by limited drug availability, toxicity concerns, diagnostic delays, biofilm-associated resistance, high mortality, and the lack of preventive vaccines. Addressing these challenges requires coordinated efforts in drug development, diagnostic innovation, and global health strategies to improve patient outcomes.

3. Mechanisms of Antifungal Resistance

Antifungal resistance is a complex and multifaceted process involving diverse molecular, genetic, and physiological adaptations that enable fungi to survive antifungal exposure. These mechanisms may occur individually or in combination, often resulting in multidrug-resistant phenotypes that significantly complicate treatment. **Figure 1** shows several mechanisms of antifungal resistance.

3.1 Drug Target Alterations

Drug target modification is a primary mechanism of antifungal resistance in which structural or quantitative changes in the target enzyme reduce drug binding and efficacy. In azole resistance, mutations in the *ERG11* gene alter the structure of lanosterol 14 α -demethylase, decreasing drug affinity, while overexpression of *ERG11* increases enzyme levels to counteract inhibition.^[82] Similarly, resistance to echinocandins arises from mutations in *FKS1* or *FKS2*, which encode subunits of β -1,3-glucan synthase, thereby reducing drug sensitivity.^[83] In the case of polyenes, alterations in membrane sterol composition, particularly reduced ergosterol content, limit drug binding and impair antifungal activity.^[84]

3.2 Efflux Pump Overexpression

Efflux pump overexpression reduces intracellular antifungal drug concentrations by actively transporting drugs out of the cell. This mechanism is primarily mediated by ATP-binding cassette (ABC) transporters, such as *Cdr1p* and *Cdr2p*, and major facilitator superfamily (MFS) transporters like *Mdr1p*.^[85] These pumps can expel a wide range of antifungal agents, particularly azoles, leading to multidrug resistance.^[86] Their expression is tightly regulated by transcription factors, including Tac1 and Mrr1, which become upregulated under antifungal stress, further enhancing resistance.^[87]

3.3 Biofilm-Mediated Resistance

Biofilm formation significantly enhances antifungal resistance by creating a protective microenvironment for fungal cells. The extracellular matrix, composed of polysaccharides, proteins, and extracellular DNA, acts as a barrier that limits drug penetration.^[88] Within biofilms, cells exhibit altered metabolic states, often entering slow-growing or dormant phases that reduce susceptibility to antifungal agents.^[89] Additionally, the presence of persister cells, a highly tolerant subpopulation, contributes to treatment failure and infection recurrence.^[90] Biofilm-associated gene expression further reinforces resistance mechanisms.

3.4 Stress Response Pathways

Fungi activate conserved stress response pathways to survive antifungal exposure and hostile host environments. The Hsp90 molecular chaperone plays a central role by stabilizing key regulatory proteins involved in resistance and stress adaptation.^[91] Calcineurin signaling, a calcium-dependent pathway,

regulates cell wall integrity, ion balance, and survival under antifungal stress.^[92] Additionally, the protein kinase C (PKC)–MAPK pathway responds to cell wall damage by inducing compensatory mechanisms such as increased chitin synthesis.^[93] Oxidative stress response systems, including enzymes like catalase and superoxide dismutase, help neutralize reactive oxygen species, further enhancing survival.^[94]

3.5 Genetic Adaptability and Genome Plasticity

Fungal pathogens exhibit remarkable genetic flexibility, allowing rapid adaptation to antifungal pressure. Aneuploidy, or variation in chromosome number, can increase the copy number of resistance-related genes, enhancing survival.^[95] Point mutations accumulate under selective pressure, while genomic instability accelerates the emergence of resistant variants.^[96] Loss of heterozygosity events can expose recessive resistance traits, particularly in diploid species.^[97] Additionally, transposable elements contribute to genomic rearrangements and altered gene expression, further promoting adaptability.^[98]

3.6 Metabolic and Pathway Rewiring

Fungi can reprogram their metabolic pathways to bypass the inhibitory effects of antifungal drugs.^[99] Alterations in the ergosterol biosynthesis pathway may lead to the accumulation of alternative sterols that partially compensate for ergosterol depletion.^[100] Changes in carbon metabolism and nutrient utilization can influence drug susceptibility by modifying cellular physiology.^[101] Furthermore, mitochondrial dysfunction or adaptation can affect energy production and stress responses, indirectly contributing to antifungal resistance.^[102]

3.7 Cell Wall Remodeling

Cell wall remodeling is a key adaptive response that enhances resistance, particularly against echinocandins.^[103] Fungi can increase chitin synthesis to compensate for the loss of β -glucan, thereby maintaining cell wall integrity despite drug exposure.^[104] Alterations in the composition and organization of cell wall polysaccharides can reduce drug accessibility and effectiveness.^[105] These structural changes not only protect the fungal cell but also contribute to immune evasion.

3.8 Epigenetic and Regulatory Mechanisms

Epigenetic modifications play an important role in regulating antifungal resistance without altering the underlying DNA sequence.^[106] Chromatin remodeling through histone modifications can influence the expression of resistance-associated genes.^[107] Additionally, RNA-mediated regulatory mechanisms, including non-coding RNAs, may modulate stress responses and drug resistance pathways.^[108] These reversible changes allow fungi to rapidly adapt to environmental stressors, including antifungal agents.

3.9 Intrinsic and Acquired Resistance

Antifungal resistance can be either intrinsic or acquired. Intrinsic resistance refers to the natural insensitivity of certain fungal species to specific antifungal agents, such as resistance of *Candida krusei* to fluconazole.^[109] In contrast, acquired resistance develops through genetic mutations, gene overexpression, or adaptive responses following exposure to antifungal drugs.^[110] This type of resistance is particularly concerning in clinical settings, where prolonged antifungal use creates strong selective pressure for resistant strains.

Overall, antifungal resistance arises from an intricate interplay of multiple mechanisms, often acting synergistically within the same organism. This complexity underscores the urgent need for novel therapeutic strategies targeting multiple pathways to effectively combat resistant fungal infections.

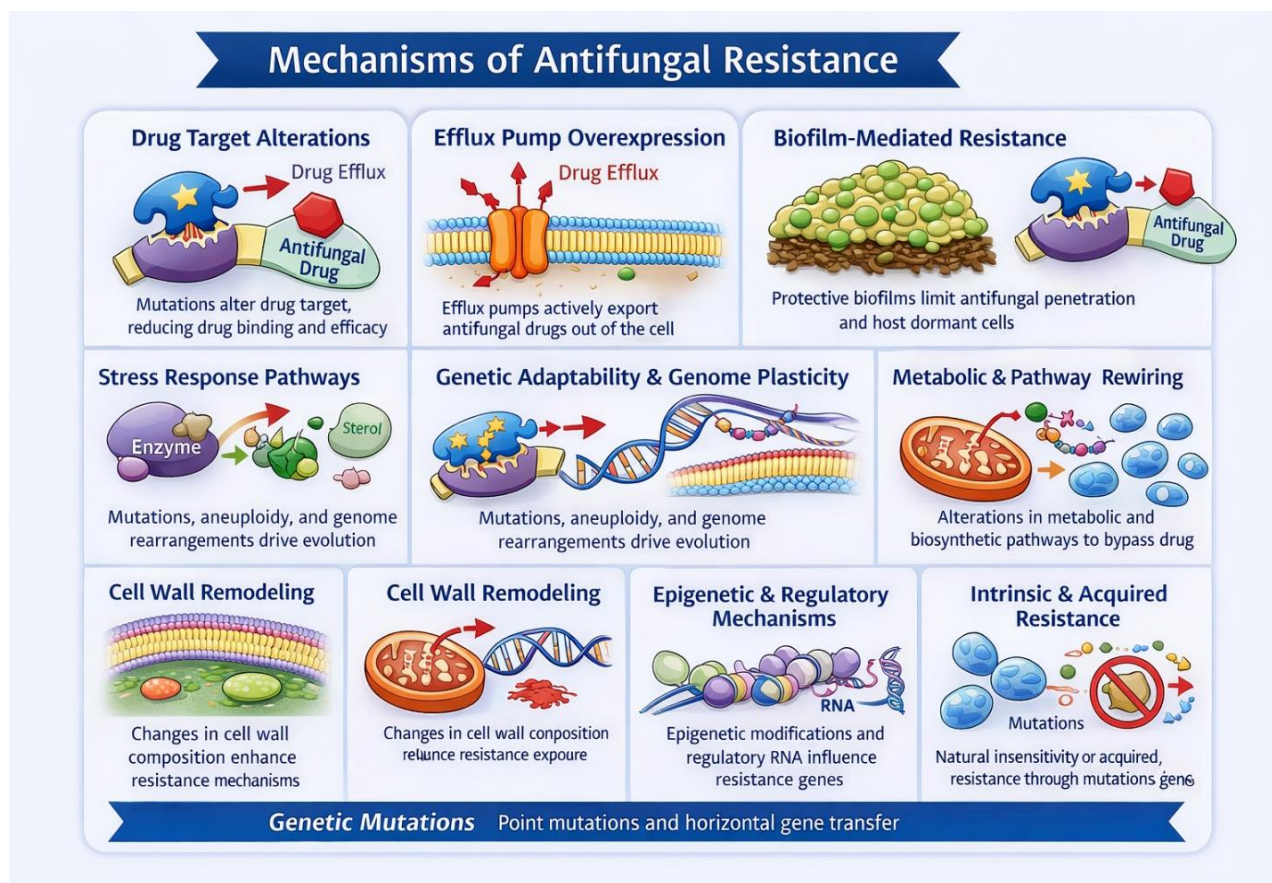


Figure 1: Schematic representation of the major mechanisms of antifungal resistance.

4. Host-Pathogen Interaction and Virulence Mechanisms

The ability of fungal pathogens to establish infection and cause disease depends on a complex interplay between host defenses and fungal virulence factors. These mechanisms enable fungi to colonize host tissues, evade immune responses, and adapt to diverse host environments, ultimately determining the severity and outcome of infection. **Figure 2** represents the key pathways of host-pathogen interactions during fungal infection.

4.1 Adhesion and Invasion

Adhesion to host cells is a critical initial step in fungal pathogenesis, allowing pathogens to colonize epithelial and mucosal surfaces. Fungi express a variety of surface adhesins, such as agglutinin-like sequence (Als) proteins in *Candida* species, which facilitate binding to host cell receptors, extracellular matrix components, and medical devices.^[111] Following adhesion, fungi can invade host tissues through both active penetration and induced endocytosis.^[112] Active penetration involves the formation of hyphal structures that mechanically breach host cell barriers, while induced endocytosis relies on host cell uptake mechanisms triggered by fungal surface proteins.^[113] These processes enable dissemination into deeper tissues and the bloodstream.

4.2 Enzyme Secretion

Fungal pathogens secrete a range of hydrolytic enzymes that contribute to tissue invasion and nutrient acquisition.^[114] Among these, proteases degrade host proteins, including structural and immune-related proteins, thereby facilitating tissue penetration and immune evasion.^[115] Phospholipases disrupt host cell membranes by hydrolyzing phospholipids, promoting cell lysis and invasion.^[116] Other enzymes, such as lipases and hemolysins, assist in nutrient acquisition and further enhance virulence.^[117] The coordinated secretion of these enzymes plays a central role in host tissue damage and pathogen survival.^[118]

4.3 Immune Evasion Strategies

Successful fungal pathogens have evolved sophisticated mechanisms to evade or modulate host immune responses. These include masking of pathogen-associated molecular patterns (PAMPs), such as β -glucans, to avoid recognition by immune receptors like Dectin-1.^[119] Some fungi produce a polysaccharide capsule, as seen in *Cryptococcus neoformans*, which inhibits phagocytosis and suppresses immune activation.^[120] Additionally, fungi can survive within phagocytic cells by resisting oxidative stress and preventing phagolysosome maturation.^[121] Modulation of host cytokine responses and interference with antigen presentation further contribute to immune evasion, allowing persistent infection.^[122]

4.4 Morphological Switching (Yeast ↔ Hyphae)

Morphological plasticity is a key virulence trait in many fungal pathogens, particularly in *Candida albicans*. The ability to switch between yeast and filamentous (hyphal or pseudohyphal) forms enables adaptation to different host environments.^[123] The yeast form is typically associated with dissemination through the bloodstream, while the hyphal form is linked to tissue invasion and biofilm formation.^[124] This transition is regulated by environmental cues such as temperature, pH, and nutrient availability, as well as complex signaling pathways within the fungal cell.^[125] Morphological switching

enhances pathogenicity by promoting adhesion, invasion, and resistance to host immune defenses.

In summary, fungal virulence is driven by coordinated mechanisms involving adhesion, enzymatic degradation of host tissues, immune evasion, and morphological adaptability. These strategies not only facilitate infection but also complicate treatment, highlighting the need for targeted therapeutic approaches that disrupt host–pathogen interactions.

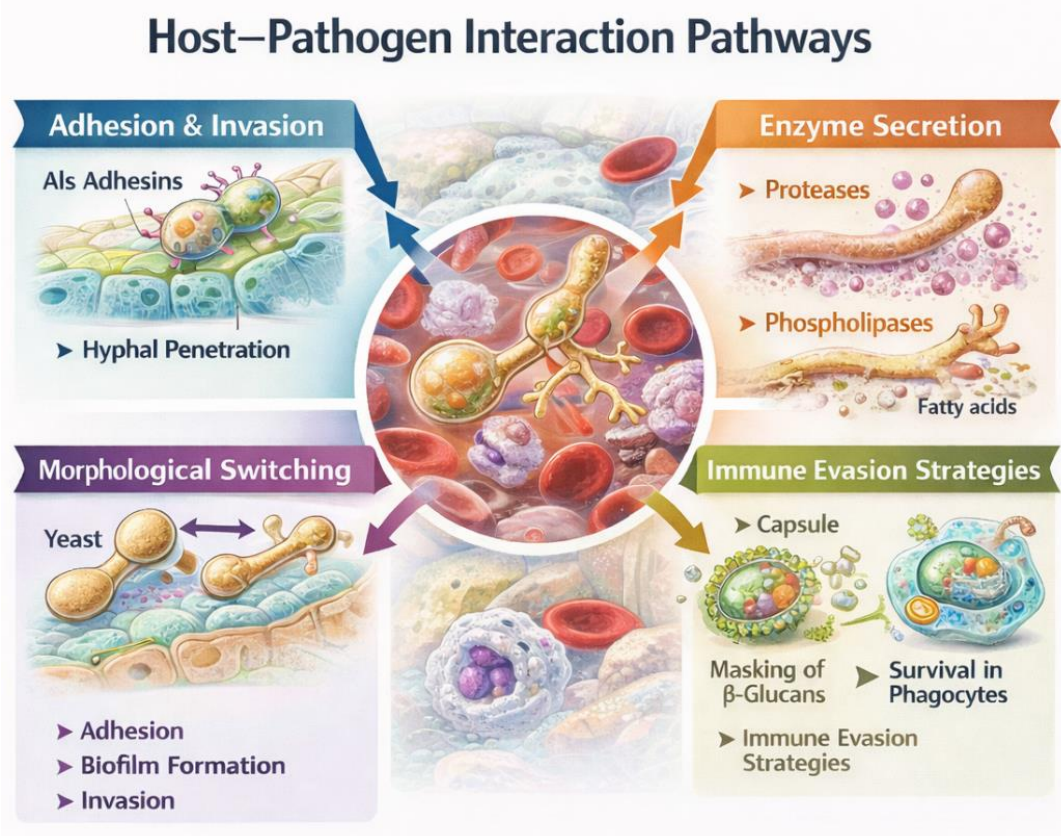


Figure 2: Host–pathogen interaction pathways in fungal infections.

5. Novel Antifungal Strategies

The growing burden of antifungal resistance, coupled with the limited efficacy and toxicity of conventional therapies, has accelerated the development of innovative and multidisciplinary antifungal strategies. These emerging approaches aim not only to improve therapeutic outcomes but also to overcome resistance, enhance drug delivery, and target previously unexplored fungal pathways. **Figure 3** represents an integrated overview of novel antifungal strategies used to combat fungal infections.

5.1 New Antifungal Agents

The antifungal drug pipeline has expanded in recent years with the development of novel agents targeting unique fungal pathways. Ibrexafungerp, a triterpenoid glucan synthase inhibitor, represents a new class with oral bioavailability and activity against echinocandin-

resistant strains.^[126] Fosmanogepix targets the Gwt1 enzyme involved in glycosylphosphatidylinositol (GPI)-anchor biosynthesis, disrupting fungal cell wall integrity and virulence.^[127] Olorofim, belonging to the orotomide class, inhibits dihydroorotate dehydrogenase in the pyrimidine biosynthesis pathway, offering a novel mechanism distinct from existing antifungals.^[128] Additional emerging agents include rezafungin (a next-generation echinocandin with extended half-life), oteseconazole (a highly selective azole), and encochleated amphotericin B formulations designed to reduce toxicity and improve delivery.^[129] **Table 2** represents a comprehensive overview of novel and emerging antifungal agents currently under development. It highlights their mechanisms of action, the fungal pathogens they target, and their respective stages in the research or clinical pipeline.

Table 2: Novel and emerging antifungal agents in development, their mechanisms of action, targeted fungi, and current stage of research or clinical development.

Agent / Class	Mechanism of Action	Target Fungi	Development Stage / Notes	Ref
Ibrexafungerp	Glucan synthase inhibitor	<i>Candida</i> spp., <i>Aspergillus</i> spp.	FDA-approved for vulvovaginal candidiasis; trials for invasive infections	[130,131]
Fosmanogepix (APX001)	Gwt1 enzyme inhibitor (cell wall synthesis)	<i>Candida</i> , <i>Aspergillus</i> , rare molds	Phase 2–3; broad-spectrum, including resistant strains	[131,132]
Olorofim (F901318)	DHODH inhibitor (pyrimidine biosynthesis)	<i>Aspergillus</i> , <i>Scedosporium</i> , <i>Lomentospora</i>	Phase 2–3; effective against azole-resistant molds	[133,134]
Rezafungin	Echinocandin (β -1,3-D-glucan synthase)	<i>Candida</i> , <i>Aspergillus</i>	Phase 3; long-acting, once-weekly dosing	[135,136]
VT-1598	Fungal CYP51 inhibitor (azole-like)	<i>Candida</i> , <i>Cryptococcus</i> , <i>Coccidioides</i>	Phase 1–2; improved selectivity and reduced toxicity	[137,138]
Nikkomyacin Z	Chitin synthase inhibitor	<i>Coccidioides</i> , <i>Candida</i>	Preclinical / early clinical; potential for combination therapy	[139,140]
MGCD290	Histone deacetylase inhibitor (synergistic with azoles)	<i>Candida</i> , <i>Aspergillus</i>	Phase 2; potentiates azole efficacy	[141,142]
T-2307	Mitochondrial function disruptor	<i>Candida</i> , <i>Cryptococcus</i>	Preclinical / early clinical; active against resistant strains	[143,144]
VT-1129	CYP51 inhibitor (next-generation azole)	<i>Cryptococcus</i> , <i>Candida</i>	Phase 2; improved oral bioavailability and safety	[145,146]
VT-1161 (Oteseconazole)	CYP51 inhibitor	<i>Candida</i> spp., vulvovaginal candidiasis	Phase 2–3; highly selective, lower drug interactions	[147,148]
ASP2397 (VL-2397)	Siderophore-mediated uptake, inhibits fungal growth	<i>Aspergillus</i> , <i>Candida</i>	Phase 2; novel uptake mechanism, active against resistant fungi	[149,150]
Aureobasidin A derivatives	Inhibits inositol phosphorylceramide synthase	<i>Candida</i> , <i>Cryptococcus</i>	Preclinical; novel cell wall target	[151,152]
SCY-078 (Ibrexafungerp analogues)	Glucan synthase inhibition	<i>Candida</i> , <i>Aspergillus</i>	Preclinical/clinical; oral formulation	[126,153]
Olorofim analogues	DHODH inhibition	Rare molds, resistant <i>Aspergillus</i>	Preclinical; improved potency and spectrum	[133,154]
Repurposed drugs (e.g., Sertraline, Tamoxifen)	Multiple mechanisms: disrupt membrane, inhibit ergosterol, affect stress response	<i>Cryptococcus</i> , <i>Candida</i>	Preclinical / off-label; adjunct therapy for resistant infections	[155,156]
Natural product derivatives (e.g., Turbinmicin, Enfumafungin derivatives)	Novel targets: biofilm inhibition, cell wall disruption	<i>Candida</i> , multidrug-resistant fungi	Preclinical; potent activity, low toxicity	[157,158]
APX001 analogues	Gwt1 inhibitors with improved pharmacokinetics	Broad-spectrum fungi	Preclinical; optimized for oral and IV use	[159,160]
Olorofim + combination therapies	DHODH inhibition + other antifungals	Resistant molds	Preclinical; synergistic effects observed	[128,161]
New echinocandin derivatives (CD101, Rezafungin analogues)	β -1,3-D-glucan synthase	<i>Candida</i> , <i>Aspergillus</i>	Phase 2–3; extended half-life, better dosing flexibility	[129,135]

5.2 Nanotechnology-Based Approaches

Nanotechnology has introduced promising strategies to enhance antifungal therapy through improved drug delivery systems. Nanoparticles, including liposomes, polymeric nanoparticles, solid lipid nanoparticles, and metallic nanoparticles (such as silver and gold), enable targeted delivery of antifungal agents to infection sites.^[162,163] These systems enhance drug solubility, stability, and bioavailability while minimizing systemic toxicity. Functionalized nanoparticles can also penetrate biofilms and deliver drugs in a controlled and sustained manner.^[164] Additionally, some nanoparticles exhibit intrinsic antifungal activity through mechanisms such as reactive oxygen species generation and membrane disruption.^[165]

5.3 Antifungal Peptides

Antifungal peptides, also known as antimicrobial peptides (AMPs), are emerging as potent alternatives to conventional antifungal drugs. These peptides typically exert their effects by disrupting fungal cell membranes, leading to leakage of intracellular contents and cell death.^[166] Some peptides also interfere with intracellular targets, including nucleic acids and protein synthesis.^[167] Natural sources include human defensins, histatins, and plant-derived peptides, while synthetic and engineered peptides are being developed to improve stability and specificity.^[168] Their broad-spectrum activity and reduced likelihood of resistance development make them highly promising candidates.

5.4 Immunotherapy

Immunotherapeutic approaches aim to enhance the host immune response against fungal pathogens. Monoclonal antibodies targeting fungal cell wall components, virulence factors, or toxins can improve pathogen clearance and reduce disease severity.^[169] Cytokine-based therapies, such as interferon- γ and granulocyte-macrophage colony-stimulating factor (GM-CSF), can boost immune cell activity and improve antifungal defense, particularly in immunocompromised patients.^[170] Other approaches include adoptive T-cell therapy and dendritic cell-based vaccines, which aim to restore or enhance antifungal immunity.^[171]

5.5 Combination Therapy

Combination antifungal therapy involves the use of two or more agents to achieve synergistic effects, improve efficacy, and reduce the likelihood of resistance development. Common strategies include combining azoles with echinocandins or polyenes, as well as pairing antifungals with non-antifungal agents such as calcineurin inhibitors or efflux pump inhibitors.^[172] These combinations can target multiple pathways simultaneously, disrupt resistance mechanisms, and enhance fungal killing.^[172] Combination therapy is particularly valuable in treating invasive and multidrug-resistant infections.

5.6 Natural Products and Phytochemicals

Natural products remain a rich source of novel antifungal compounds with diverse mechanisms of action. Plant-derived compounds such as alkaloids, flavonoids, terpenoids, and phenolics exhibit antifungal activity by disrupting cell membranes, inhibiting enzyme function, and interfering with biofilm formation.^[173,174] Essential oils from plants, including components like thymol, eugenol, and carvacrol, have demonstrated strong antifungal effects.^[175,176] In addition, secondary metabolites from microorganisms, marine organisms, and endophytic fungi are being explored for their therapeutic potential.^[177] These compounds often exhibit synergistic effects when combined with conventional antifungals.

5.7 Targeting Fungal Virulence Factors

An emerging strategy involves targeting fungal virulence rather than viability, thereby reducing selective pressure for resistance. This includes inhibition of adhesion, biofilm formation, enzyme secretion, and morphological switching.^[178] Quorum sensing inhibitors and biofilm-disrupting agents are being investigated to interfere with fungal communication and pathogenicity without directly killing the organism.^[179]

5.8 Gene- and RNA-Based Therapeutics

Advances in molecular biology have enabled the development of gene-targeting strategies such as RNA interference (RNAi) and CRISPR-Cas systems. These approaches can selectively silence or disrupt genes essential for fungal survival, virulence, or resistance.^[180] Although still largely experimental, they offer high specificity and the potential for personalized antifungal therapy.

5.9 Drug Repurposing Strategies

Drug repurposing involves the use of existing non-antifungal drugs with newly identified antifungal properties. Compounds such as statins, antipsychotics, anticancer drugs, and immunomodulators have shown activity against fungal pathogens.^[181] This approach reduces development time and cost, as safety profiles are already established, and can be particularly useful in rapidly addressing emerging resistance.

5.10 Microbiome-Based Approaches

The role of the human microbiome in controlling fungal colonization has gained increasing attention. Probiotics and microbiome-modulating therapies can inhibit fungal overgrowth by competing for nutrients and producing antifungal metabolites.^[182] Restoration of microbial balance may serve as a preventive or adjunctive strategy in fungal infections.^[183]

5.11 Photodynamic and Physical Therapies

Photodynamic therapy (PDT) utilizes photosensitizing agents activated by specific wavelengths of light to generate reactive oxygen species that kill fungal cells.^[184] Other physical approaches, such as cold plasma and

ultrasound-assisted therapy, are also being explored for their antifungal effects, particularly in superficial and biofilm-associated infections.^[185]

So, novel antifungal strategies encompass a wide range of innovative approaches, from new drug development to

advanced delivery systems and host-directed therapies. These strategies aim to overcome the limitations of current antifungal treatments and provide more effective, targeted, and sustainable solutions for managing emerging fungal infections.

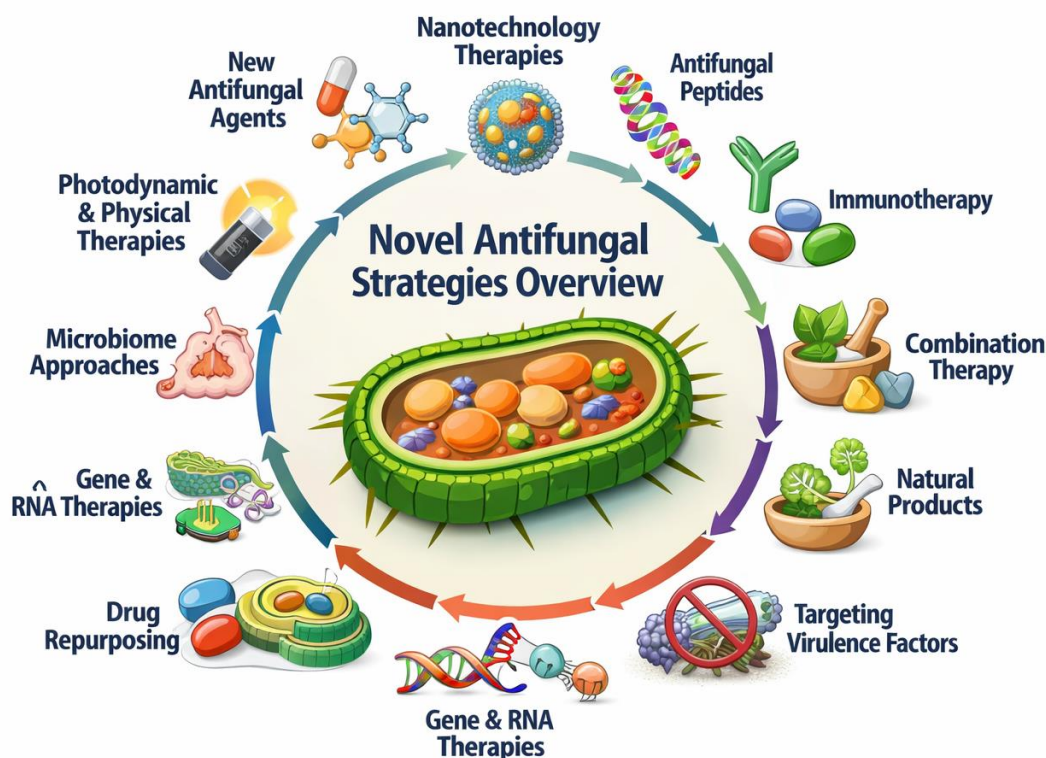


Figure 3: Novel antifungal strategies overview.

6. Diagnostic Advances

Timely and accurate diagnosis of fungal infections is critical for effective clinical management, yet remains challenging due to nonspecific symptoms and limitations of conventional methods. Recent advances in diagnostic technologies have significantly improved the speed, sensitivity, and specificity of fungal detection, enabling earlier intervention and better patient outcomes.

6.1 PCR-Based Detection

Polymerase chain reaction (PCR)-based techniques have revolutionized fungal diagnostics by allowing rapid and highly sensitive detection of fungal DNA directly from clinical samples.^[186] These methods can identify pathogens at the species level, even in cases where cultures are negative or slow-growing. Real-time PCR (qPCR) and multiplex PCR assays enable simultaneous detection of multiple fungal species, making them particularly useful in invasive infections.^[187] Additionally, PCR-based methods can detect antifungal resistance genes, providing valuable information for targeted therapy.^[188] However, challenges such as standardization, cost, and risk of contamination still limit widespread clinical implementation.

6.2 MALDI-TOF Mass Spectrometry

Matrix-Assisted Laser Desorption/Ionization Time-of-Flight (MALDI-TOF) mass spectrometry has emerged as a powerful tool for the rapid identification of fungal pathogens. This technique analyzes protein spectra from microbial cells and compares them with reference databases to accurately identify species within minutes.^[189] MALDI-TOF is particularly effective for identifying *Candida* and other yeast species, and advancements are expanding its application to filamentous fungi.^[190] Its advantages include speed, cost-effectiveness over time, and high accuracy, although its performance depends on the quality and completeness of reference databases.^[191]

6.3 Biomarker-Based Detection

Biomarker assays provide non-culture-based methods for detecting fungal infections by identifying specific fungal cell wall components or metabolites in body fluids.^[192] Among the most widely used biomarkers is β -D-glucan, a component of the fungal cell wall found in many pathogenic fungi, which serves as a broad indicator of invasive fungal infection.^[193] Galactomannan, another important biomarker, is specifically associated with *Aspergillus* infections and is commonly used for early

diagnosis of invasive aspergillosis.^[194] These assays offer rapid results and are particularly useful for screening high-risk patients; however, they may produce false positives or negatives under certain clinical conditions, necessitating careful interpretation alongside clinical findings.^[195]

DISCUSSION

The emergence of multidrug-resistant fungal pathogens represents one of the most pressing challenges in modern infectious disease management. Over the past decade, there has been a notable shift in the epidemiology of invasive fungal infections, with species such as *Candida auris*, non-*albicans Candida*, azole-resistant *Aspergillus fumigatus*, and *Cryptococcus gattii* gaining prominence.^[196] These pathogens are associated with higher mortality rates, increased healthcare costs, and prolonged hospital stays, particularly in immunocompromised populations, including patients undergoing chemotherapy, organ transplantation, or long-term corticosteroid therapy.^[197] The increasing prevalence of these infections highlights the inadequacy of current antifungal options and underscores the urgent need for novel therapeutic and preventive strategies.

A central concern in the management of emerging fungal infections is the growing antifungal resistance. Resistance arises through multiple mechanisms, including drug target alterations, efflux pump overexpression, biofilm-mediated tolerance, activation of stress response pathways, and genomic plasticity.^[198,199] For instance, mutations in the *ERG11* gene in *Candida* species reduce azole binding, while alterations in FKS genes confer echinocandin resistance.^[200] Biofilms further complicate treatment, providing a protective niche where fungi can survive high drug concentrations.^[77] These resistance mechanisms often coexist within the same pathogen, producing multidrug-resistant phenotypes that are increasingly difficult to eradicate. Resistance not only limits treatment options but also drives the need for higher drug doses, combination therapies, and prolonged treatment courses, which in turn increase toxicity risks and healthcare burden.

The limitations of current antifungal agents contribute significantly to the clinical challenges. Only three major classes—azoles, echinocandins, and polyenes—are widely used, with each exhibiting inherent limitations.^[201] Polyenes, such as amphotericin B, remain highly effective but are associated with nephrotoxicity and infusion-related adverse effects.^[72] Azoles are prone to drug–drug interactions and hepatotoxicity, while echinocandins, although generally well-tolerated, have limited oral bioavailability and reduced efficacy against certain resistant strains.^[73] Moreover, the lack of antifungal vaccines or prophylactic immunization strategies leaves at-risk populations highly susceptible to invasive infections, further emphasizing the dependence on pharmacological interventions.

Advances in diagnostic technologies offer some hope in improving clinical outcomes by enabling early detection. Molecular techniques, such as PCR-based assays, allow for rapid species-level identification and the detection of resistance-associated genes.^[202] MALDI-TOF mass spectrometry has accelerated fungal identification with high accuracy and reduced turnaround time.^[190] Biomarker-based assays, including β -D-glucan and galactomannan detection, provide additional tools for early diagnosis of systemic infections.^[203] Despite these advances, diagnostic accessibility and standardization remain significant barriers, particularly in resource-limited settings, leading to delayed treatment initiation and increased mortality.

The urgent need for innovative therapies has driven research into novel antifungal strategies. New agents such as ibrexafungerp, fosmanogepix, and olorofim target unique fungal pathways and exhibit activity against multidrug-resistant strains.^[204] Nanotechnology-based delivery systems enhance drug solubility, targeted delivery, and biofilm penetration while reducing systemic toxicity.^[205] Antifungal peptides offer broad-spectrum activity through membrane disruption, and immunotherapeutic approaches, including monoclonal antibodies and cytokine therapy, aim to augment host defenses.^[206] Additionally, combination therapy, natural products, and phytochemicals provide complementary approaches that may improve efficacy and reduce resistance development.^[207] Importantly, strategies targeting fungal virulence factors, rather than viability alone, represent a promising direction to reduce selective pressure for resistance.^[208]

Beyond clinical management, surveillance and a One Health perspective are critical for controlling emerging fungal threats. Environmental reservoirs, agricultural use of azoles, and climate-driven shifts in fungal distribution all contribute to the emergence of resistant strains.^[209] Coordinated global surveillance systems, integrated with genomic epidemiology, can track resistance trends and guide therapeutic strategies.^[210] The One Health approach, linking human, animal, and environmental health, is essential to understanding and mitigating the factors driving fungal emergence and resistance.

Despite these advances, several research gaps persist. Identification of new drug targets, development of safe and effective vaccines, standardization of rapid diagnostics, and scalable surveillance systems remain priorities.^[211] Additionally, understanding the molecular mechanisms of biofilm tolerance, stress response, and genetic adaptability will inform the design of more effective antifungal therapies.^[212] Interdisciplinary collaboration between mycologists, clinicians, pharmacologists, and public health experts is essential to address these gaps and translate laboratory findings into clinical practice.

Therefore, the current landscape of emerging fungal pathogens is characterized by increasing resistance, high morbidity and mortality, and limited treatment options. Innovative therapies, rapid diagnostics, global surveillance, and integrated preventive strategies are essential to address this escalating threat. Continued research and investment in antifungal drug discovery, immunotherapy, and One Health initiatives will be critical in mitigating the clinical and public health impact of emerging fungal infections. Addressing these challenges proactively will not only improve patient outcomes but also reduce the global burden of fungal diseases.

Future Perspectives and Research Gaps

Despite progress in understanding fungal biology and improving therapies, major gaps still limit control of emerging fungal infections. Addressing these challenges requires coordinated efforts across research, clinical practice, and global health systems.

A key priority is the discovery of novel antifungal drug targets. Current treatments focus on a few pathways, such as ergosterol synthesis and cell wall biosynthesis, increasing resistance risks.^[213] Future research should explore fungal-specific pathways, including virulence factors, mitochondrial functions, and unique metabolic processes.^[214] Advances in genomics and systems biology can accelerate the identification of these targets and support safer, more effective drug development.

Vaccine development remains a significant challenge. Fungi are complex eukaryotes, making it difficult to identify safe, immunogenic targets, especially in immunocompromised patients who are most at risk.^[215] Current approaches—including subunit, conjugate, and live-attenuated vaccines—face hurdles in safety, efficacy, and durability, which must be overcome before widespread clinical application.^[216]

Surveillance systems are critical for monitoring resistance and emerging infections. Existing data are often fragmented and region-specific. Global networks with standardized diagnostics, reporting, and genomic integration can enable early detection of resistant strains, track multidrug-resistant outbreaks, and guide public health strategies.^[217]

A One Health approach is essential, as fungal infections are influenced by human, animal, and environmental factors. Agricultural antifungal use and climate-driven changes contribute to resistant environmental strains.^[218] Collaboration across medical, veterinary, and environmental sectors is needed to regulate antifungal use, monitor reservoirs, and implement sustainable control strategies.^[219]

So, future research should focus on expanding drug targets, overcoming vaccine barriers, strengthening surveillance, and adopting One Health strategies. These

efforts are crucial to prevent, manage, and reduce the global impact of emerging fungal pathogens.

CONCLUSION

Emerging fungal pathogens represent a growing global health threat, causing severe infections with high morbidity and mortality, particularly among immunocompromised individuals. The rapid rise of multidrug-resistant species, such as *Candida auris* and azole-resistant *Aspergillus fumigatus*, underscores the urgent antifungal resistance crisis. Conventional therapies are increasingly limited by toxicity, biofilm-associated tolerance, and evolving resistance mechanisms.

Addressing this challenge requires a multifaceted approach that includes the development of novel antifungal agents, advanced drug delivery systems, immunotherapies, and combination strategies. Early and accurate diagnostics, coupled with global surveillance and One Health initiatives, are equally essential to monitor and contain resistant strains. Continued investment in innovative therapies and preventive measures, including vaccines and natural product-based interventions, will be critical to mitigating the threat posed by emerging fungal pathogens and improving clinical outcomes worldwide.

Acknowledgement

None

Conflict of Interest

Nil

REFERENCES

1. Anandani G, Bhise M, Agarwal A. Invasive fungal infections and the management in immunocompromised conditions. *J Fam Med Prim care*. 2025; 14(7): 2643-2652. doi: 10.4103/jfmpc.jfmpc_1582_24
2. Branda F, Petrosillo N, Ceccarelli G, et al. Antifungal Agents in the 21st Century: Advances, Challenges, and Future Perspectives. *Infect Dis Rep*. 2025; 17(4). doi: 10.3390/idr17040091
3. Kim JS, Cha H, Bahn YS. Comprehensive Overview of *Candida auris*: An Emerging Multidrug-Resistant Fungal Pathogen. *J Microbiol Biotechnol*. 2024; 34(7): 1365-1375. doi: 10.4014/jmb.2404.04040
4. Berger S, El Chazli Y, Babu AF, Coste AT. Azole Resistance in *Aspergillus fumigatus*: A Consequence of Antifungal Use in Agriculture? *Front Microbiol*. 2017; 8: 1024. doi: 10.3389/fmicb.2017.01024
5. Zhao Y, Ye L, Zhao F, et al. Cryptococcus neoformans, a global threat to human health. *Infect Dis poverty*. 2023; 12(1): 20. doi: 10.1186/s40249-023-01073-4
6. George ME, Gaitor TT, Cluck DB, Henao-Martínez AF, Sells NR, Chastain DB. The impact

- of climate change on the epidemiology of fungal infections: implications for diagnosis, treatment, and public health strategies. *Ther Adv Infect Dis*. 2025; 12: 20499361251313840. doi: 10.1177/20499361251313841
7. Revie NM, Iyer KR, Robbins N, Cowen LE. Antifungal drug resistance: evolution, mechanisms and impact. *Curr Opin Microbiol*. 2018; 45: 70-76. doi: 10.1016/j.mib.2018.02.005
 8. Wolfgruber S, Salmanton-García J, Kuate MPN, Hoenigl M, Brunelli JGP. Antifungal pipeline: New tools for the treatment of mycoses. *Rev Iberoam Micol*. 2024; 41(4): 68-78. doi: <https://doi.org/10.1016/j.riam.2024.11.001>
 9. Suneja M, Beekmann SE, Dhaliwal G, Miller AC, Polgreen PM. Diagnostic delays in infectious diseases. *Diagnosis (Berlin, Ger)*. 2022; 9(3): 332-339. doi: 10.1515/dx-2021-0092
 10. Rather MA, Gupta K, Mandal M. Microbial biofilm: formation, architecture, antibiotic resistance, and control strategies. *Brazilian J Microbiol [publication Brazilian Soc Microbiol]*. 2021; 52(4): 1701-1718. doi: 10.1007/s42770-021-00624-x
 11. Banawas SS. Epidemiology of Fungal Bloodstream Infections and Antifungal Susceptibility in a Tertiary Care Hospital in Riyadh, Saudi Arabia: A Rare Candida Co-Infection Case. *Pathog (Basel, Switzerland)*. 2025; 14(12). doi: 10.3390/pathogens14121221
 12. Sanyaolu A, Okorie C, Marinkovic A, et al. Candida auris: An Overview of the Emerging Drug-Resistant Fungal Infection. *Infect Chemother*. 2022; 54(2): 236-246. doi: 10.3947/ic.2022.0008
 13. Pandey N, Gupta MK, Paul P, Tilak R. Necessity to identify candida species accurately with minimum inhibitory concentration determination in each case of bloodstream infections. *J Infect Public Health*. 2020; 13(5): 753-758. doi: <https://doi.org/10.1016/j.jiph.2019.12.002>
 14. Gómez-Gaviria M, Ramírez-Sotelo U, Mora-Montes HM. Non-albicans Candida Species: Immune Response, Evasion Mechanisms, and New Plant-Derived Alternative Therapies. *J fungi (Basel, Switzerland)*. 2022; 9(1). doi: 10.3390/jof9010011
 15. Sugui JA, Kwon-Chung KJ, Juvvadi PR, Latgé JP, Steinbach WJ. Aspergillus fumigatus and related species. *Cold Spring Harb Perspect Med*. 2014; 5(2): a019786. doi: 10.1101/cshperspect.a019786
 16. Dabholkar A, Pandit S, Devkota R, et al. Role of the osaA Gene in Aspergillus fumigatus Development, Secondary Metabolism and Virulence. *J fungi (Basel, Switzerland)*. 2024; 10(2). doi: 10.3390/jof10020103
 17. Cao bw, Wang F, Yu S, et al. Prevalence of Azole-Resistant Aspergillus fumigatus is Highly Associated with Azole Fungicide Residues in the Fields. *Environ Sci Technol*. 2021; 55. doi: 10.1021/acs.est.0c03958
 18. Hsu TH, Huang PY, Fan YC, Sun PL. Azole Resistance and cyp51A Mutation of Aspergillus fumigatus in a Tertiary Referral Hospital in Taiwan. *J fungi (Basel, Switzerland)*. 2022; 8(9). doi: 10.3390/jof8090908
 19. Rafique A, Sharmin S, Raj A, Mohiuddin AL, Mahmud MI Al, Bin Md Omer H. Comparative overview of Aspergillus fumigatus, A. flavus, and A. niger: Pathogenicity, resistance, and public health significance. *J Infect Public Health*. 2026; 19(2): 103070. doi: <https://doi.org/10.1016/j.jiph.2025.103070>
 20. Nikhil A, Choudhury S, Bhatia M, et al. Antifungal Susceptibility Profile of Aspergillus Strains Isolated From the Lower Respiratory Tract in Eastern Indian Patients: A Hospital-Based Study. *Microbiologyopen*. 2025; 14(6): e70136. doi: 10.1002/mbo3.70136
 21. Diniz-Lima I, Fonseca LM da, Silva-Junior EB da, et al. Cryptococcus: History, Epidemiology and Immune Evasion. *Appl Sci*. 2022; 12(14). doi: 10.3390/app12147086
 22. Mm H, Chakraborty S, Aa K. Cryptococcal Meningoencephalitis in an Immunosuppressed Patient with Chronic Lymphocytic Leukemia. 2020; 16(2): 87-89.
 23. Gibson JF, Johnston SA. Immunity to Cryptococcus neoformans and C. gattii during cryptococcosis. *Fungal Genet Biol*. 2015; 78: 76-86. doi: 10.1016/j.fgb.2014.11.006
 24. Manogaran Y, Ramasamy P. Cryptococcus neoformans and Cryptococcus gattii species in urban tropics: A comprehensive review of pathogenicity and resistance. *The Microbe*. 2025; 8: 100493. doi: <https://doi.org/10.1016/j.microb.2025.100493>
 25. Kavitha J, Sivakrishnan S, Lakshmisree S, Srinivasan S. Mucormycosis - Life threatening invasive fungal disease. 12(3): 440-447.
 26. Kumar R, Misra AK, Dutta S, Gupta A, Kumar B, Charan J. A systematic review of mucormycosis cases in COVID-19: Is it an unholy trilogy of COVID-19, diabetes mellitus, and corticosteroids? *J Fam Med Prim care*. 2022; 11(6): 2573-2580. doi: 10.4103/jfmpc.jfmpc_1934_21
 27. Ledoux M pierre, Dicop E, Sabou M, et al. Fusarium , Scedosporium and Other Rare Mold Invasive Infections: Over Twenty-Five-Year Experience of a European. Published online 2024.
 28. Marinelli T, Kim HY, Halliday CL, et al. Fusarium species, Scedosporium species, and Lomentospora prolificans: A systematic review to inform the World Health Organization priority list of fungal pathogens. *Med Mycol*. 2024; 62(6). doi: 10.1093/mmy/myad128
 29. Li LX, Yoon H. Dematiaceous Molds. *Infect Dis Clin North Am*. 2025; 39(1): 75-92. doi: 10.1016/j.idc.2024.11.006
 30. America S. Scientific letter. Published online 2023: 581-583.

31. Giusiano G. Systemic endemic mycoses: From a geographical risk to a concern expansion. *Rev Iberoam Micol.* 2025; 42(3): 87-92. doi: <https://doi.org/10.1016/j.riam.2025.07.001>
32. Sil A, Andrianopoulos A. Thermally Dimorphic Human Fungal Pathogens--Polyphyletic Pathogens with a Convergent Pathogenicity Trait. *Cold Spring Harb Perspect Med.* 2014; 5(8): a019794. doi: [10.1101/cshperspect.a019794](https://doi.org/10.1101/cshperspect.a019794)
33. Kourti M, Roilides E. Invasive Trichosporonosis in Neonates and Pediatric Patients with Malignancies or Hematologic Disorders. *Pathog (Basel, Switzerland).* 2022; 11(2). doi: [10.3390/pathogens11020242](https://doi.org/10.3390/pathogens11020242)
34. Vazquez JA, Whitaker L, Zubovskaia A. Invasive Candidiasis in the Intensive Care Unit : Where Are We Now ? Published online 2025.
35. Bir R, Chatterjee K, Rai A, et al. Candida Species Isolated From ICU Bloodstream Infections: Molecular Epidemiology, Antifungal Resistance, and Virulence Profiling. *Cureus.* 2025; 17(10): e94898. doi: [10.7759/cureus.94898](https://doi.org/10.7759/cureus.94898)
36. Bocci MG, Cascarano L, Capecchi G, et al. Pulmonary Aspergillosis in Immunocompromised Critically Ill Patients: Prevalence, Risk Factors, Clinical Features and Diagnosis-A Narrative Review. *J fungi (Basel, Switzerland).* 2025; 11(9). doi: [10.3390/jof11090617](https://doi.org/10.3390/jof11090617)
37. Antonia AL, Alspaugh JA. Cryptococcus : Emerging host risk factors for infection. Published online 2025: 1-10. doi: [10.1371/journal.ppat.1013602](https://doi.org/10.1371/journal.ppat.1013602)
38. Mourad A, Perfect JR. Present and Future Therapy of Cryptococcus Infections. *J fungi (Basel, Switzerland).* 2018; 4(3). doi: [10.3390/jof4030079](https://doi.org/10.3390/jof4030079)
39. Skiada A, Drogari-apiranthitou M, Pavleas I, Daikou E, Petrikkos G. Global Cutaneous Mucormycosis : A Systematic Review. Published online 2022: 1-17.
40. Morrissey CO, Kim HY, Garnham K, et al. Mucorales: A systematic review to inform the World Health Organization priority list of fungal pathogens. *Med Mycol.* 2024; 62(6). doi: [10.1093/mmy/myad130](https://doi.org/10.1093/mmy/myad130)
41. Ferreira J, Plinio W, Makris G, Sandoval-denis M, Hagen F, Crous PW. Fusarioid keratitis and other superficial infections : A 10-years prospective study from Northeastern Brazil. Published online 2024: 1-22. doi: [10.1371/journal.pntd.0012247](https://doi.org/10.1371/journal.pntd.0012247)
42. Valeri S, Alvarez-moreno C, Pape P Le, et al. A One Health Perspective to Recognize Fusarium as Important in Clinical Practice.
43. Seidel D, Hassler A, Salmanton-garcía J, et al. International Journal of Infectious Diseases Invasive Scedosporium spp . and Lomentospora proli fi cans infections in pediatric patients : Analysis of 55 cases from FungiScope 1 and the literature. 2020; 92: 114-122. doi: [10.1016/j.ijid.2019.12.017](https://doi.org/10.1016/j.ijid.2019.12.017)
44. Liu W, Feng R, Jiang H. Management of pulmonary Scedosporium apiospermum infection by thoracoscopic surgery in an immunocompetent woman. *J Int Med Res.* 2020; 48(7): 300060520931620. doi: [10.1177/0300060520931620](https://doi.org/10.1177/0300060520931620)
45. Kondratjeva J, Pressanti C, Reynolds BS, et al. Multifocal cutaneous phaeohyphomycosis caused by Exophiala spinifera with clinical resolution in an immunocompromised cat. Published online 2023. doi: [10.1177/20551169231164610](https://doi.org/10.1177/20551169231164610)
46. Usuda D, Higashikawa T, Hotchi Y, et al. Exophiala dermatitidis. *World J Clin cases.* 2021; 9(27): 7963-7972. doi: [10.12998/wjcc.v9.i27.7963](https://doi.org/10.12998/wjcc.v9.i27.7963)
47. Duvenage L, Salie S, Keeton R, et al. The pathogenesis of experimental Emergomycosis in mice. Published online 2024: 1-19. doi: [10.1371/journal.pntd.0011850](https://doi.org/10.1371/journal.pntd.0011850)
48. Rooms I, Mugisha P, Gambichler T, et al. Disseminated Emergomycosis in a Person with HIV Infection, Uganda. 2019; 25(9): 1750-1751.
49. Afonso P, Cardoso L, Soares AS, Matos M, Quintas H, Coelho AC. Talaromyces marneffeii Outside Endemic Regions: An Overlooked Mycosis Under a One-Health Lens. *Acta Microbiol Hell.* 2025; 70(2). doi: [10.3390/amh70020025](https://doi.org/10.3390/amh70020025)
50. Qin Y, Huang X, Chen H, et al. Burden of Talaromyces marneffeii infection in people living with HIV/AIDS in Asia during ART era: a systematic review and meta-analysis. *BMC Infect Dis.* 2020; 20(1): 551. doi: [10.1186/s12879-020-05260-8](https://doi.org/10.1186/s12879-020-05260-8)
51. McDonald EG, Afshar A, Assiri B, et al. Pneumocystis jirovecii pneumonia in people living with HIV : a review.
52. Iriart X, Bouar M Le, Kamar N, Berry A. Pneumocystis Pneumonia in Solid-Organ Transplant Recipients. *J fungi (Basel, Switzerland).* 2015; 1(3): 293-331. doi: [10.3390/jof1030293](https://doi.org/10.3390/jof1030293)
53. Zhang H, Zhang J. Trichosporon asahii: emerging challenges in pathogenesis and drug resistance. *Future Microbiol.* 2025; 20(4): 333-343. doi: [10.1080/17460913.2025.2457858](https://doi.org/10.1080/17460913.2025.2457858)
54. Santos FAG, Leite-Andrade MC, Vasconcelos MARA, et al. Trichosporon Inkin Fungemia Case Report: Clinical and Laboratory Management. *Future Microbiol.* 2022; 17(2): 81-87. doi: [10.2217/fmb-2021-0017](https://doi.org/10.2217/fmb-2021-0017)
55. Al-Shaarani AAQA, Pecoraro L. A review of pathogenic airborne fungi and bacteria: unveiling occurrence, sources, and profound human health implication. *Front Microbiol.* 2024; 15: 1428415. doi: [10.3389/fmicb.2024.1428415](https://doi.org/10.3389/fmicb.2024.1428415)
56. Niyazi D, Daskalova N, Micheva I, Stoeva T. Cladosporium cladosporioides Fungemia in a Patient with Non-Hodgkin Lymphoma: An Extremely Rare Case and Review of the Literature. *Reports.* 2026; 9(1). doi: [10.3390/reports9010060](https://doi.org/10.3390/reports9010060)
57. Evans J, Wang A, Elewski B. Successful Treatment of Paecilomyces lilacinus Onychomycosis with Efinaconazole and Tavaborole. *Ski Appendage*

- Disord.* 2016; 1: 169-171. doi: 10.1159/000443773
58. Calendino S, Kenna J, Patel H, et al. Aggressive Purpurocillium lilacinum Kerato-Endophthalmitis in a Diabetic Contact Lens Wearer Culminating in Enucleation: A Case Report and Review of the Literature. *J fungi (Basel, Switzerland)*. 2025; 11(11). doi: 10.3390/jof11110789
 59. McBride JA, Sterkel AK, Matkovic E, Broman AT, Gibbons-Burgener SN, Gauthier GM. Clinical Manifestations and Outcomes in Immunocompetent and Immunocompromised Patients With Blastomycosis. *Clin Infect Dis an Off Publ Infect Dis Soc Am.* 2021; 72(9): 1594-1602. doi: 10.1093/cid/ciaa276
 60. Linder KA, Kauffman CA, Miceli MH. Blastomycosis: A Review of Mycological and Clinical Aspects. *J fungi (Basel, Switzerland)*. 2023; 9(1). doi: 10.3390/jof9010117
 61. Cáceres DH, Gómez BL, Tobón ÁM, et al. Tackling Histoplasmosis Infection in People Living with HIV from Latin America: From Diagnostic Strategy to Public Health Solutions. *J Fungi.* 2023; 9(5). doi: 10.3390/jof9050558
 62. Allan R, Domingues S, Regina M, Oliveira F De. Disseminated Histoplasmosis in People Living With HIV: What Are the Care Costs for Brazil? *Value Heal Reg Issues.* 2025; 50: 101166. doi: 10.1016/j.vhri.2025.101166
 63. Hahn RC, Rodrigues AM, Portella P, et al. Clinical and epidemiological features of paracoccidioidomycosis due to Paracoccidioides lutzii. 2019; 7: 1-13.
 64. de Almeida JNJ, Peçanha-Pietrobon PM, Colombo AL. Paracoccidioidomycosis in Immunocompromised Patients: A Literature Review. *J fungi (Basel, Switzerland)*. 2018; 5(1). doi: 10.3390/jof5010002
 65. Rakotoarisaona MF, Andrianarison M, Sendrasoa FA, Rasamoelina T, Ramarozatovo LS, Rapelanoro Rabenja F. Cutaneous disseminated sporotrichosis in an immunocompetent farmer. *Med Mycol Case Rep.* 2024; 43: 100626. doi: https://doi.org/10.1016/j.mmcr.2023.100626
 66. Queiroz-Telles F, Buccheri R, Benard G. Sporotrichosis In Immunocompromised Hosts. *J fungi (Basel, Switzerland)*. 2019; 5(1). doi: 10.3390/jof5010008
 67. Hattab Z, Ben Lasfar N, Abid M, et al. Alternaria alternata infection causing rhinosinusitis and orbital involvement in an immunocompetent patient. *New microbes new Infect.* 2019; 32: 100561. doi: 10.1016/j.nmni.2019.100561
 68. Paccoud O, Vignier N, Boui M, et al. Invasive Rhinosinusitis Caused by Alternaria infectoria in a Patient with Autosomal Recessive CARD9 Deficiency and a Review of the Literature. *J fungi (Basel, Switzerland)*. 2022; 8(5). doi: 10.3390/jof8050446
 69. Van Rhijn N, White PL. Antifungal treatment strategies and their impact on resistance development in clinical settings. *J Antimicrob Chemother.* 2025; 80(12): 3208-3226. doi: 10.1093/jac/dkaf382
 70. Lu X, Zhou J, Ming Y, et al. Next-generation antifungal drugs: Mechanisms, efficacy, and clinical prospects. *Acta Pharm Sin B.* 2025; 15(8): 3852-3887. doi: https://doi.org/10.1016/j.apsb.2025.06.013
 71. Li Y, Liu Y, Jiang Y, et al. New antifungal strategies and drug development against WHO critical priority fungal pathogens. *Front Cell Infect Microbiol.* 2025; 15: 1662442. doi: 10.3389/fcimb.2025.1662442
 72. Carolus H, Pierson S, Lagrou K, Van Dijck P. Amphotericin B and Other Polyenes-Discovery, Clinical Use, Mode of Action and Drug Resistance. *J fungi (Basel, Switzerland)*. 2020; 6(4). doi: 10.3390/jof6040321
 73. Rakhshan A, Rahmati Kamel B, Saffaei A, Tavakoli-Ardakani M. Hepatotoxicity Induced by Azole Antifungal Agents: A Review Study. *Iran J Pharm Res IJPR.* 2023; 22(1): e130336. doi: 10.5812/ijpr-130336
 74. Rosam K, Steixner S, Bauer A. Expert Review of Molecular Diagnostics Non-conventional diagnostic methods for invasive fungal infections. *Expert Rev Mol Diagn.* 2025; 25(7): 313-328. doi: 10.1080/14737159.2025.2509026
 75. Dinnes J, Deeks JJ, Adriano A, et al. Rapid, point-of-care antigen and molecular-based tests for diagnosis of SARS-CoV-2 infection. *Cochrane database Syst Rev.* 2020; 8(8): CD013705. doi: 10.1002/14651858.CD013705
 76. Zhao A, Sun J, Liu Y. Understanding bacterial biofilms: From definition to treatment strategies. *Front Cell Infect Microbiol.* 2023; 13: 1137947. doi: 10.3389/fcimb.2023.1137947
 77. Asokan S, Pandey RK, Jalil MA, et al. Biofilm associated infections on medical devices: Pathogenesis, diagnostic challenges, and control strategies. *The Microbe.* 2026; 11: 100712. doi: https://doi.org/10.1016/j.microb.2026.100712
 78. Denning DW. Global incidence and mortality of severe fungal disease. *Lancet Infect Dis.* 2024; 24(7): e428-e438. doi: 10.1016/S1473-3099(23)00692-8
 79. Akalu TY, Clements ACA, Gebreyohannes EA, Xu Z, Bai L, Alene KA. Risk factors for diagnosis and treatment delay among patients with multidrug-resistant tuberculosis in Hunan Province, China. *BMC Infect Dis.* 2024; 24(1): 159. doi: 10.1186/s12879-024-09036-2
 80. Inácio MM, Moreira ALE, Cruz-Leite VRM, et al. Fungal Vaccine Development: State of the Art and Perspectives Using Immunoinformatics. *J fungi (Basel, Switzerland)*. 2023; 9(6). doi: 10.3390/jof9060633
 81. Rivera A, Lodge J, Xue C. Harnessing the Immune Response to Fungal Pathogens for Vaccine Development. *Annu Rev Microbiol.* 2022; 76: 703-

726. doi: 10.1146/annurev-micro-041020-111511
82. Graham DO, Wilson RK, Ruma YN, Keniya M V, Tyndall JDA, Monk BC. Structural Insights into the Azole Resistance of the *Candida albicans* Darlington Strain Using *Saccharomyces cerevisiae* Lanosterol 14 α -Demethylase as a Surrogate. *J fungi (Basel, Switzerland)*. 2021; 7(11). doi: 10.3390/jof7110897
 83. Zajac C, Scott NE, Kline S, Erayil SE, Selmecki A. Hotspot gene conversion between FKS1 and FKS2 in echinocandin resistant *Candida glabrata* serial isolates. *npj Antimicrob Resist*. Published online 2025. doi: 10.1038/s44259-025-00102-6
 84. Haro-Reyes T, Díaz-Peralta L, Galván-Hernández A, Rodríguez-López A, Rodríguez-Fragoso L, Ortega-Blake I. Polyene Antibiotics Physical Chemistry and Their Effect on Lipid Membranes; Impacting Biological Processes and Medical Applications. *Membranes (Basel)*. 2022; 12(7). doi: 10.3390/membranes12070681
 85. Banerjee A, Pata J, Sharma S, Monk BC, Falson P, Prasad R. Directed Mutational Strategies Reveal Drug Binding and Transport by the MDR Transporters of *Candida albicans*. *J fungi (Basel, Switzerland)*. 2021; 7(2). doi: 10.3390/jof7020068
 86. Holmes AR, Cardno TS, Strouse JJ, et al. Targeting efflux pumps to overcome antifungal drug resistance. *Future Med Chem*. 2016; 8(12): 1485-1501. doi: 10.4155/fmc-2016-0050
 87. Chang W, Liu J, Zhang M, et al. triphenylphosphonium cation. *Nat Commun*. (2018): 1-12. doi: 10.1038/s41467-018-07633-9
 88. Gunn JS, Bakaletz LO, Wozniak DJ. What's on the Outside Matters: The Role of the Extracellular Polymeric Substance of Gram-negative Biofilms in Evading Host Immunity and as a Target for Therapeutic Intervention. *J Biol Chem*. 2016; 291(24): 12538-12546. doi: 10.1074/jbc.R115.707547
 89. Debta P, Sahu BK, Patra SK, Debta FM, Mishra E, Panda SK. Overcoming *Candida* biofilm resistance: targeting persister cells with probiotic-derived metabolites. *Front Antibiot*. 2026; 5: 1767028. doi: 10.3389/frabi.2026.1767028
 90. Wuyts J, Van Dijck P, Holtappels M. Fungal persister cells: The basis for recalcitrant infections? *PLoS Pathog*. 2018; 14(10): e1007301. doi: 10.1371/journal.ppat.1007301
 91. Aguilar-Rodríguez J, Jakobson CM, Jarosz DF. The Hsp90 Molecular Chaperone as a Global Modifier of the Genotype-Phenotype-Fitness Map: An Evolutionary Perspective. *J Mol Biol*. 2024; 436(23): 168846. doi: <https://doi.org/10.1016/j.jmb.2024.168846>
 92. Liu S, Hou Y, Liu W, Lu C, Wang W, Sun S. Components of the calcium-calcineurin signaling pathway in fungal cells and their potential as antifungal targets. *Eukaryot Cell*. 2015; 14(4): 324-334. doi: 10.1128/EC.00271-14
 93. Manfiolli AO, Dos Reis TF, de Assis LJ, et al. Mitogen activated protein kinases (MAPK) and protein phosphatases are involved in *Aspergillus fumigatus* adhesion and biofilm formation. *Cell Surf (Amsterdam, Netherlands)*. 2018; 1: 43-56. doi: 10.1016/j.tcsw.2018.03.002
 94. Jomova K, Raptova R, Alomar SY, et al. Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. *Arch Toxicol*. 2023; 97(10): 2499-2574. doi: 10.1007/s00204-023-03562-9
 95. Ravichandran MC, Fink S, Clarke MN, Hofer FC, Campbell CS. Genetic interactions between specific chromosome copy number alterations dictate complex aneuploidy patterns. *Genes Dev*. 2018; 32(23-24): 1485-1498. doi: 10.1101/gad.319400.118
 96. Ambrosio AD, Bressan D, Ferracci E, et al. Increased genomic instability and reshaping of tissue microenvironment underlie oncogenic properties of *Arid1a* mutations. 2024; 4435(March). doi: 10.1126/sciadv.adh4435
 97. Smukowski Heil C. Loss of Heterozygosity and Its Importance in Evolution. *J Mol Evol*. 2023; 91(3): 369-377. doi: 10.1007/s00239-022-10088-8
 98. Bhat A, Ghatage T, Bhan S, et al. Role of Transposable Elements in Genome Stability: Implications for Health and Disease. *Int J Mol Sci*. 2022; 23(14). doi: 10.3390/ijms23147802
 99. Parente-Rocha JA, Bailão AM, Amaral AC, et al. Antifungal Resistance, Metabolic Routes as Drug Targets, and New Antifungal Agents: An Overview about Endemic Dimorphic Fungi. *Mediators Inflamm*. 2017; 2017: 9870679. doi: 10.1155/2017/9870679
 100. Navarro-Mendoza MI, Pérez-Arques C, Parker J, Xu Z, Kelly S, Heitman J. Alternative ergosterol biosynthetic pathways confer antifungal drug resistance in the human pathogens within the *Mucor* species complex. *MBio*. 2024; 15(8): e0166124. doi: 10.1128/mbio.01661-24
 101. Koh EI, Oluoch PO, Ruecker N, et al. Chemical-genetic interaction mapping links carbon metabolism and cell wall structure to tuberculosis drug efficacy. *Proc Natl Acad Sci U S A*. 2022; 119(15): e2201632119. doi: 10.1073/pnas.2201632119
 102. Alves R, Gourlay CW. Editorial: Mitochondrial function and dysfunction in pathogenic fungi. *Front Physiol*. 2024; 15: 1506684. doi: 10.3389/fphys.2024.1506684
 103. Davis FA, Singh K, Krampen JM, et al. Bacterial metabolites induce cell wall remodeling, antifungal resistance, and immune recognition of commensal fungi. *Curr Biol*. 2026; 36(3): 674-691.e8. doi: <https://doi.org/10.1016/j.cub.2025.12.029>
 104. Brain L, Bleackley M, Doblin MS, Anderson M. Fungal Chitin Synthases: Structure, Function, and Regulation. *J fungi (Basel, Switzerland)*. 2025; 11(11). doi: 10.3390/jof11110796
 105. Visan AI, Cristescu R. Polysaccharide-Based

- Coatings as Drug Delivery Systems. *Pharmaceutics*. 2023; 15(9). doi: 10.3390/pharmaceutics15092227
106. Patra S, Raney M, Pareek A, Kaur R. Epigenetic Regulation of Antifungal Drug Resistance. *J fungi (Basel, Switzerland)*. 2022; 8(8). doi: 10.3390/jof8080875
 107. Kang H, Fan T, Wu J, Zhu Y, Shen WH. Histone modification and chromatin remodeling in plant response to pathogens. *Front Plant Sci*. 2022; Volume 13-2022. doi: 10.3389/fpls.2022.986940
 108. Ghadyani Nejhad L, Sohani M, Ghandforoush NA, et al. Non-coding RNAs: Emerging contributors to chemoresistance in chronic myeloid leukemia. *Leuk Res reports*. 2025; 23: 100513. doi: 10.1016/j.lrr.2025.100513
 109. Akbar Z, Aamir M, Saleem Z, et al. Antifungal Resistance Among Candida Species: Diagnostic and Clinical Challenges in Specialized Cancer Care Hospital of Lahore. *J Clin Lab Anal*. 2025; 39(9): e70022. doi: 10.1002/jcla.70022
 110. Kadariswantiningsih IN, Empitu MA, Santosa TI, Alimu Y. Antifungal resistance: Emerging mechanisms and implications (Review). *Mol Med Rep*. 2025; 32(3). doi: 10.3892/mmr.2025.13612
 111. Hoyer LL, Cota E. Candida albicans Agglutinin-Like Sequence (Als) Family Vignettes: A Review of Als Protein Structure and Function. *Front Microbiol*. 2016; Volume 7-2016. doi: 10.3389/fmicb.2016.00280
 112. Sheppard DC, Filler SG. Host cell invasion by medically important fungi. *Cold Spring Harb Perspect Med*. 2014; 5(1): a019687. doi: 10.1101/cshperspect.a019687
 113. Tamai R, Kiyoura Y. Candida Infections: The Role of Saliva in Oral Health—A Narrative Review. *Microorganisms*. 2025; 13(4). doi: 10.3390/microorganisms13040717
 114. Corrêa-Junior D, Zamith-Miranda D, Frases S, Nosanchuk JD. From the Ground to the Clinic: The Evolution and Adaptation of Fungi. *J fungi (Basel, Switzerland)*. 2025; 12(1). doi: 10.3390/jof12010008
 115. Xia Q, Liu X, Huang H. Host proteases: key regulators in viral infection and therapeutic targeting. 2025; (September). doi: 10.3389/fimmu.2025.1671173
 116. Ali U, Lu S, Fadlalla T, et al. The functions of phospholipases and their hydrolysis products in plant growth, development and stress responses. *Prog Lipid Res*. 2022; 86: 101158. doi: 10.1016/j.plipres.2022.101158
 117. Kuhn HW, Lasseter AG, Adams PP, et al. BB0562 is a nutritional virulence determinant with lipase activity important for *Borrelia burgdorferi* infection and survival in fatty acid deficient environments. *PLoS Pathog*. 2021; 17(8): e1009869. doi: 10.1371/journal.ppat.1009869
 118. Ramírez-Larrota JS, Eckhard U. An Introduction to Bacterial Biofilms and Their Proteases, and Their Roles in Host Infection and Immune Evasion. *Biomolecules*. 2022; 12(2). doi: 10.3390/biom12020306
 119. Pradhan A, Avelar GM, Bain JM, et al. Non-canonical signalling mediates changes in fungal cell wall PAMPs that drive immune evasion. *Nat Commun*. (2019): 1-14. doi: 10.1038/s41467-019-13298-9
 120. Decote-Ricardo D, LaRocque-de-Freitas IF, Rocha JDB, et al. Immunomodulatory Role of Capsular Polysaccharides Constituents of *Cryptococcus neoformans*. *Front Med*. 2019; 6: 129. doi: 10.3389/fmed.2019.00129
 121. Ruiz-Baca E, Adame-Soto PJ, Alba-Fierro CA, et al. Response to Oxidative Stress in *Sporothrix schenckii*. *J Fungi*. 2025; 11(6). doi: 10.3390/jof11060440
 122. Tarasova O, Petrou A, Ivanov SM, Geronikaki A, Poroikov V. Viral Factors in Modulation of Host Immune Response: A Route to Novel Antiviral Agents and New Therapeutic Approaches. *Int J Mol Sci*. 2024; 25(17). doi: 10.3390/ijms25179408
 123. Chow EWL, Pang LM, Wang Y. From Jekyll to Hyde: The Yeast-Hyphal Transition of *Candida albicans*. *Pathog (Basel, Switzerland)*. 2021; 10(7). doi: 10.3390/pathogens10070859
 124. Amann V, Kissmann AK, Firacative C, Rosenau F. Biofilm-Associated Candidiasis: Pathogenesis, Prevalence, Challenges and Therapeutic Options. *Pharmaceutics*. 2025; 18(4). doi: 10.3390/ph18040460
 125. Ibe C, Munro CA. Fungal Cell Wall Proteins and Signaling Pathways Form a Cytoprotective Network to Combat Stresses. *J fungi (Basel, Switzerland)*. 2021; 7(9). doi: 10.3390/jof7090739
 126. Jallow S, Govender NP. Ibrexafungerp: A First-in-Class Oral Triterpenoid Glucan Synthase Inhibitor. *J fungi (Basel, Switzerland)*. 2021; 7(3). doi: 10.3390/jof7030163
 127. Shaw KJ, Ibrahim AS. Fosmanogepix: A Review of the First-in-Class Broad Spectrum Agent for the Treatment of Invasive Fungal Infections. *J fungi (Basel, Switzerland)*. 2020; 6(4). doi: 10.3390/jof6040239
 128. Vanbiervliet Y, Van Nieuwenhuysse T, Aerts R, Lagrou K, Spriet I, Maertens J. Review of the novel antifungal drug olorofim (F901318). *BMC Infect Dis*. 2024; 24(1): 1256. doi: 10.1186/s12879-024-10143-3
 129. Sofjan AK, Mitchell A, Shah DN, et al. Rezafungin (CD101), a next-generation echinocandin: A systematic literature review and assessment of possible place in therapy. *J Glob Antimicrob Resist*. 2018; 14: 58-64. doi: 10.1016/j.jgar.2018.02.013
 130. He R, Lin F, Yu B, Huang L. Efficacy and safety of ibrexafungerp in the treatment of vulvovaginal candidiasis: A meta-analysis of randomized controlled trials. *Heliyon*. 2024; 10(8): e28776. doi: https://doi.org/10.1016/j.heliyon.2024.e28776
 131. El Ayoubi LW, Allaw F, Moussa E, Kanj SS.

- Ibrexafungerp: A narrative overview. *Curr Res Microb Sci.* 2024; 6: 100245. doi: 10.1016/j.crmicr.2024.100245
132. Trzoss M, Covell JA, Kapoor M, et al. Synthesis of analogs of the Gwt1 inhibitor manogepix (APX001A) and in vitro evaluation against *Cryptococcus* spp. *Bioorg Med Chem Lett.* 2019; 29(23): 126713. doi: <https://doi.org/10.1016/j.bmcl.2019.126713>
 133. Maertens JA, Thompson GR, Spec A, et al. Olorofim for the treatment of invasive fungal diseases in patients with few or no therapeutic options: a single-arm, open-label, phase 2b study. *Lancet Infect Dis.* 2025; 25(11): 1177-1188. doi: [https://doi.org/10.1016/S1473-3099\(25\)00224-5](https://doi.org/10.1016/S1473-3099(25)00224-5)
 134. Seyedmousavi S, Chang YC, Law D, Birch M, Rex JH, Kwon-Chung KJ. Efficacy of Olorofim (F901318) against *Aspergillus fumigatus*, *A. nidulans*, and *A. tanneri* in Murine Models of Profound Neutropenia and Chronic Granulomatous Disease. *Antimicrob Agents Chemother.* 2019; 63(6). doi: 10.1128/AAC.00129-19
 135. Luo H, Zang Y, Zhang D. Rezafungin: A β -(1,3)-D-glucan synthase inhibitor as antifungal drug derived from Echinocandin B0. In: 2026: 207-224. doi: 10.1016/B978-0-443-33885-4.00014-7
 136. Andes D, Brüggemann RJ, Flanagan S, et al. The distinctive pharmacokinetic profile of rezafungin, a long-acting echinocandin developed in the era of modern pharmacometrics. *J Antimicrob Chemother.* 2025; 80(1): 18-28. doi: 10.1093/jac/dkae415
 137. Gu K, Spitz R, Hammett E, et al. Safety and pharmacokinetics of antifungal agent VT-1598 and its primary metabolite, VT-11134, in healthy adult subjects: phase 1, first-in-human, randomized, double-blind, placebo-controlled study of single-ascending oral doses of VT-1598. *Med Mycol.* 2024; 62(4). doi: 10.1093/mmy/myae032
 138. Wiederhold NP, Patterson HP, Tran BH, Yates CM, Schotzinger RJ, Garvey EP. Fungal-specific Cyp51 inhibitor VT-1598 demonstrates in vitro activity against *Candida* and *Cryptococcus* species, endemic fungi, including *Coccidioides* species, *Aspergillus* species and *Rhizopus arrhizus*. *J Antimicrob Chemother.* 2018; 73(2): 404-408. doi: 10.1093/jac/dkx410
 139. Wu Y, Zhang M, Yang Y, et al. Structures and mechanism of chitin synthase and its inhibition by antifungal drug Nikkomycin Z. Published online 2022: 4-7. doi: 10.1038/s41421-022-00495-y
 140. Larwood DJ. Nikkomycin Z-Ready to Meet the Promise? *J fungi (Basel, Switzerland).* 2020; 6(4). doi: 10.3390/jof6040261
 141. Garnaud C, Champeboux M, Maubon D, Cornet M. Histone Deacetylases and Their Inhibition in *Candida* Species. 2016; 7(August): 1-10. doi: 10.3389/fmicb.2016.01238
 142. Li C, Tu J, Han G, Liu N, Sheng C. Heat shock protein 90 (Hsp90)/Histone deacetylase (HDAC) dual inhibitors for the treatment of azoles-resistant *Candida albicans*. *Eur J Med Chem.* 2022; 227: 113961. doi: <https://doi.org/10.1016/j.ejmech.2021.113961>
 143. Wiederhold NP. Review of T-2307, an Investigational Agent That Causes Collapse of Fungal Mitochondrial Membrane Potential. *J Fungi.* 2021; 7(2). doi: 10.3390/jof7020130
 144. Qin Y, Wang J. Recent Progress in Research on Mitochondrion-Targeted Antifungal Drugs: a Review. 2023; (May).
 145. Puumala E, Fallah S, Robbins N, Cowen LE. Advancements and challenges in antifungal therapeutic development. *Clin Microbiol Rev.* 2024; 37(1): e00142-23. doi: 10.1128/cmr.00142-23
 146. Lockhart S, Iqbal N, Bolden C, et al. The Investigational Fungal Cyp51 Inhibitor VT-1129 Demonstrates Potent In Vitro Activity against *Cryptococcus neoformans* and *Cryptococcus gattii*. *Antimicrob Agents Chemother.* 2016; 60. doi: 10.1128/AAC.02770-15
 147. Author C. Oteseconazole - An advance in the treatment of Recurrent Vulvovaginal Candidiasis? 2023; 6(1).
 148. Brand SR, Sobel JD, Nyirjesy P, Ghannoum MA, Schotzinger RJ, Degenhardt TP. A Randomized Phase 2 Study of VT-1161 for the Treatment of Acute Vulvovaginal Candidiasis. *Clin Infect Dis an Off Publ Infect Dis Soc Am.* 2021; 73(7): e1518-e1524. doi: 10.1093/cid/ciaa1204
 149. Nakamura I, Ohsumi K, Takeda S, et al. ASP2397 Is a Novel Natural Compound That Exhibits Rapid and Potent Fungicidal Activity against *Aspergillus* Species through a Specific Transporter. *Antimicrob Agents Chemother.* 2019; 63(10): 10.1128/aac.02689-18. doi: 10.1128/aac.02689-18
 150. Shaw KJ. GR-2397: Review of the Novel Siderophore-like Antifungal Agent for the Treatment of Invasive Aspergillosis. *J fungi (Basel, Switzerland).* 2022; 8(9). doi: 10.3390/jof8090909
 151. Teimouri M, Shams-Ghahfarokhi M, Razzaghi-Abyaneh M. Inhibitory effects and mechanism of antifungal action of the natural cyclic depsipeptide, aureobasidin A against *Cryptococcus neoformans*. *Bioorg Med Chem Lett.* 2021; 41. doi: 10.1016/j.bmcl.2021.128013
 152. Zhen C, Lu H, Jiang Y. Novel Promising Antifungal Target Proteins for Conquering Invasive Fungal Infections. *Front Microbiol.* 2022; Volume 13-2022. doi: 10.3389/fmicb.2022.911322
 153. Rivero-Menendez O, Soto-Debran JC, Cuenca-Estrella M, Alastruey-Izquierdo A. In Vitro Activity of Ibrexafungerp against a Collection of Clinical Isolates of *Aspergillus*, Including Cryptic Species and Cyp51A Mutants, Using EUCAST and CLSI Methodologies. *J Fungi.* 2021; 7(3). doi: 10.3390/jof7030232
 154. Cui X, Wang L, Lü Y, Yue C. Development and research progress of anti-drug resistant fungal

- drugs. *J Infect Public Health*. 2022; 15(9): 986-1000. doi: <https://doi.org/10.1016/j.jiph.2022.08.004>
155. Peyclit L, Yousfi H, Rolain JM, Bittar F. Drug Repurposing in Medical Mycology: Identification of Compounds as Potential Antifungals to Overcome the Emergence of Multidrug-Resistant Fungi. *Pharmaceutics*. 2021; 14(5). doi: 10.3390/ph14050488
 156. Rodríguez-Cerdeira C, Eckhardt W. Therapeutic Repurposing of Sertraline: Evidence for Its Antifungal Activity from In Vitro, In Vivo, and Clinical Studies. *Microorganisms*. 2025; 13(10). doi: 10.3390/microorganisms13102334
 157. Kemkuignou BM, Haahr M, Maleckis M, Ding L. Discovery and biosynthesis of antifungal microbial secondary metabolites. *Microbiol Mol Biol Rev*. 2026; 90(1): e00075-25. doi: 10.1128/mmbr.00075-25
 158. Ganeshkumar A, Muthuselvam M, Lima PMN de, Rajaram R, Junqueira JC. Current Perspectives of Antifungal Therapy: A Special Focus on *Candida auris*. *J fungi (Basel, Switzerland)*. 2024; 10(6). doi: 10.3390/jof10060408
 159. Shaw KJ, Schell WA, Covell J, et al. *In Vitro* and *In Vivo* Evaluation of APX001A/APX001 and Other Gwt1 Inhibitors against *Cryptococcus*. *Antimicrob Agents Chemother*. 2018; 62(8): 10.1128/aac.00523-18. doi: 10.1128/aac.00523-18
 160. Viriyakosol S, Kapoor M, Okamoto S, et al. APX001 and Other Gwt1 Inhibitor Prodrugs Are Effective in Experimental *Coccidioides immitis* Pneumonia. *Antimicrob Agents Chemother*. 2019; 63(2). doi: 10.1128/AAC.01715-18
 161. Pinder C, Lebedinec R, Oliver JD, Birch M, Law D. In vitro evaluation of olorofim and antifungal combinations against *Aspergillus* and *Candida* species. *J Antimicrob Chemother*. 2025; 80(11): 3139-3149. doi: 10.1093/jac/dkaf354
 162. Islam S, Ahmed MMS, Islam MA, Hossain N, Chowdhury MA. Advances in nanoparticles in targeted drug delivery—A review. *Results in Surfaces and Interfaces*. 2025; 19: 100529. doi: <https://doi.org/10.1016/j.rsufi.2025.100529>
 163. Tarannum N, Pooja K, Jakhar S, Mavi A. Nanoparticles assisted intra and transdermic delivery of antifungal ointment: an updated review. *Discov nano*. 2024; 19(1): 11. doi: 10.1186/s11671-023-03932-3
 164. Guo Y, Mao Z, Ran F, et al. Nanotechnology-Based Drug Delivery Systems to Control Bacterial-Biofilm-Associated Lung Infections. *Pharmaceutics*. 2023; 15(11). doi: 10.3390/pharmaceutics15112582
 165. Wu Q, Cen F, Xie Y, et al. Nanoparticle-based antifungal therapies innovations mechanisms and future prospects. *PeerJ*. 2025; 13: e19199. doi: 10.7717/peerj.19199
 166. Buda De Cesare G, Cristy SA, Garsin DA, Lorenz MC. Antimicrobial Peptides: a New Frontier in Antifungal Therapy. *MBio*. 2020; 11(6). doi: 10.1128/mBio.02123-20
 167. Buyanova M, Pei D. Targeting intracellular protein-protein interactions with macrocyclic peptides. *Trends Pharmacol Sci*. 2022; 43(3): 234-248. doi: 10.1016/j.tips.2021.11.008
 168. Nagib M, Sayed AM, Korany AH, et al. Human Defensins: Structure, Function, and Potential as Therapeutic Antimicrobial Agents with Highlights Against SARS CoV-2. *Probiotics Antimicrob Proteins*. 2025; 17(3): 1563-1583. doi: 10.1007/s12602-024-10436-8
 169. Stepanyan L, Israyelyan M, Gori A, et al. Natural and Synthetic Peptides as Alternatives to Antibiotics in Intestinal Infections—A Review. *Antibiotics*. 2026; 15(1). doi: 10.3390/antibiotics15010068
 170. Dong Q, Lu J, Liu M, Wu W, Kang Y, Zhang R. The power of GM-CSF: immune regulation in the defense against *Phialophora verrucosa* infection. *Front Immunol*. 2025; 16: 1662183. doi: 10.3389/fimmu.2025.1662183
 171. Motallebzadeh Khanmiri J, Khani-Eshratabadi M, Seyedmoharrami F, et al. Innovative combinatory approaches with dendritic cell-based vaccines: bridging preclinical insights and clinical challenges. *Clin Exp Med*. 2026; 26(1): 154. doi: 10.1007/s10238-026-02056-z
 172. Toepfer S, Keniya M V, Lackner M, Monk BC. Azole Combinations and Multi-Targeting Drugs That Synergistically Inhibit *Candida auris*. *J fungi (Basel, Switzerland)*. 2024; 10(10). doi: 10.3390/jof10100698
 173. Dantas TDS, Machado JCB, Ferreira MRA, Soares LAL. Bioactive Plant Compounds as Alternatives Against Antifungal Resistance in the *Candida* Strains. *Pharmaceutics*. 2025; 17(6). doi: 10.3390/pharmaceutics17060687
 174. Huang W, Wang Y, Tian W, et al. Biosynthesis Investigations of Terpenoid, Alkaloid, and Flavonoid Antimicrobial Agents Derived from Medicinal Plants. *Antibiot (Basel, Switzerland)*. 2022; 11(10). doi: 10.3390/antibiotics11101380
 175. Moussa H, Omari B El, Chefchaou H, et al. Action of thymol, carvacrol and eugenol on *Penicillium* and *Geotrichum* isolates resistant to commercial fungicides and causing postharvest citrus decay Disease control / Moyens de lutte Action of thymol, carvacrol and eugenol on *Penicillium* and *Geotrichum* isolates resistant to commercial fungicides and causing postharvest citrus decay. *Can J Plant Pathol*. 2021; 43(1): 26-34. doi: 10.1080/07060661.2020.1767692
 176. Papantzikos V, Patakioutas G, Yfanti P. Evaluation of the Antifungal Effect of Carvacrol-Rich Essential Oils: In Vitro Study on the Phytopathogenic Fungi *Alternaria* and *Fusarium*. *Biol Life Sci Forum*. 2025; 54(1). doi: 10.3390/blsf2025054001
 177. Singh VK, Kumar A. Secondary metabolites from

- endophytic fungi: Production, methods of analysis, and diverse pharmaceutical potential. *Symbiosis*. Published online June 2023: 1-15. doi: 10.1007/s13199-023-00925-9
178. Ivanov M, Ćirić A, Stojković D. Emerging Antifungal Targets and Strategies. *Int J Mol Sci*. 2022; 23(5). doi: 10.3390/ijms23052756
 179. Singh S, Bhatia S. Quorum Sensing Inhibitors: Curbing Pathogenic Infections through Inhibition of Bacterial Communication. *Iran J Pharm Res IJPR*. 2021; 20(2): 486-514. doi: 10.22037/ijpr.2020.113470.14318
 180. Ansari RA, Rezaee Danesh Y, Castello I, Vitale A. Molecular Identification and RNA-Based Management of Fungal Plant Pathogens: From PCR to CRISPR/Cas9. *Int J Mol Sci*. 2026; 27(2). doi: 10.3390/ijms27021073
 181. E Silva DM, de Souza Lacerda L, de Souza Andrioli A, et al. Statins as Antifungal Agents: A Review on Drug Repurposing and Nanotechnology-Driven Delivery Strategies. *Fundam Clin Pharmacol*. 2025; 39(5): e70046. doi: 10.1111/fcp.70046
 182. Wu Y, Hu S, Wu C, Gu F, Yang Y. Probiotics: Potential Novel Therapeutics Against Fungal Infections. *Front Cell Infect Microbiol*. 2021; 11: 793419. doi: 10.3389/fcimb.2021.793419
 183. Agbadamashi DJ, Price CL. Novel Strategies for Preventing Fungal Infections-Outline. *Pathog (Basel, Switzerland)*. 2025; 14(2). doi: 10.3390/pathogens14020126
 184. Tanu R, Chaudhary AA, Prakash G, et al. Exploring the potential of photodynamic therapy in overcoming multidrug resistance: mechanisms, synergies, and clinical advancements in infectious diseases. *Front Cell Infect Microbiol*. 2025; Volume 15-2025. doi: 10.3389/fcimb.2025.1624036
 185. Davis CC, Dias Panariello F, Panariello B. Exploring the Efficacy of Low-Temperature Plasmas on Oral Biofilms: A Scoping Review. *Med Sci (Basel, Switzerland)*. 2025; 13(2). doi: 10.3390/medsci13020079
 186. Brown L, Cruciani M, Morton CO, et al. The Molecular Diagnosis of Invasive Fungal Diseases with a Focus on PCR. *Diagnostics*. 2025; 15(15). doi: 10.3390/diagnostics15151909
 187. Ismadi YKM, Mohamad S, Harun A. Development of multiplex real-time PCR for simultaneous detection of common fungal pathogens in invasive mycoses. *PeerJ*. 2024; 12: e18238. doi: 10.7717/peerj.18238
 188. Garcia-Effron G. Molecular Markers of Antifungal Resistance: Potential Uses in Routine Practice and Future Perspectives. *J fungi (Basel, Switzerland)*. 2021; 7(3). doi: 10.3390/jof7030197
 189. Giordano ALPL, Pontes L, Beraquet CAG, Lyra L, Schreiber AZ. Matrix-assisted laser desorption/ionisation-time of flight mass spectrometry azole susceptibility assessment in *Candida* and *Aspergillus* species. *Mem Inst Oswaldo Cruz*. 2023; 118: e220213. doi: 10.1590/0074-02760220213
 190. Lau AF. Matrix-Assisted Laser Desorption Ionization Time-of-Flight for Fungal Identification. *Clin Lab Med*. 2021; 41(2): 267-283. doi: 10.1016/j.cll.2021.03.006
 191. Czeszewska-Rosiak G, Adamczyk I, Ludwiczak A, et al. Analysis of the efficacy of MALDI-TOF MS technology in identifying microorganisms in cancer patients and oncology hospital environment. *Heliyon*. 2025; 11(2): e42015. doi: https://doi.org/10.1016/j.heliyon.2025.e42015
 192. Rivera-Agudelo L, Cáceres DH, Arango-Rincón JC, Jaramillo-Alzate JC, Zuluaga-Rodríguez A, Rúa-Giraldo ÁL. Biomarkers in the Diagnosis of Aspergillosis. *J Fungi*. 2026; 12(4). doi: 10.3390/jof12040259
 193. Morjaria S, Esther Babady N. Maximizing the diagnostic potential of 1,3 beta-D-glucan assays for invasive fungal infections. *Clin Microbiol Newsl*. 2025; 50: 24-33. doi: https://doi.org/10.1016/j.clinmicnews.2025.01.001
 194. Többen C, Cornely OA, Joisten CS, et al. Sequential serum galactomannan as outcome marker for invasive aspergillosis—An exploratory study from the FungiScope registry. *Int J Infect Dis*. 2026; 163: 108272. doi: https://doi.org/10.1016/j.ijid.2025.108272
 195. Zhao Q, Zhang C, Zhang W, Zhang S, Liu Q, Guo Y. Applications and challenges of biomarker-based predictive models in proactive health management. *Front Public Heal*. 2025; Volume 13-2025. doi: 10.3389/fpubh.2025.1633487
 196. Arastehfar A, Gabaldón T, Garcia-Rubio R, et al. Drug-Resistant Fungi: An Emerging Challenge Threatening Our Limited Antifungal Armamentarium. *Antibiot (Basel, Switzerland)*. 2020; 9(12). doi: 10.3390/antibiotics9120877
 197. Seagle EE, Williams SL, Chiller TM. Recent Trends in the Epidemiology of Fungal Infections. *Infect Dis Clin North Am*. 2021; 35(2): 237-260. doi: 10.1016/j.idc.2021.03.001
 198. Elshobary ME, Badawy NK, Ashraf Y, et al. Combating Antibiotic Resistance: Mechanisms, Multidrug-Resistant Pathogens, and Novel Therapeutic Approaches: An Updated Review. *Pharmaceuticals*. 2025; 18(3). doi: 10.3390/ph18030402
 199. Elbaiomy RG, El-Sappah AH, Guo R, et al. Antibiotic Resistance: A Genetic and Physiological Perspective. *MedComm*. 2025; 6(11): e70447. doi: 10.1002/mco2.70447
 200. Hou X, Lee A, Jiménez-Ortigosa C, Kordalewska M, Perlin DS, Zhao Y. Rapid Detection of ERG11-Associated Azole Resistance and FKS-Associated Echinocandin Resistance in *Candida auris*. *Antimicrob Agents Chemother*. 2019; 63(1). doi: 10.1128/AAC.01811-18
 201. Lakhani P, Patil A, Majumdar S. Challenges in the

- Polyene- and Azole-Based Pharmacotherapy of Ocular Fungal Infections. *J Ocul Pharmacol Ther Off J Assoc Ocul Pharmacol Ther*. 2019; 35(1): 6-22. doi: 10.1089/jop.2018.0089
202. Alsuoef EA, Alsayed AR, Zraikat MS, et al. Molecular Detection of Antibiotic Resistance Genes Using Respiratory Sample from Pneumonia Patients. *Antibiotics*. 2025; 14(5). doi: 10.3390/antibiotics14050502
 203. Forster J, Koc Ö, Koeppl MB, et al. β -1,3-d-Glucan and Galactomannan as Biomarkers for the Detection of Invasive Geotrichum and Magnusiomyces Infections: a Retrospective Evaluation. *J Clin Microbiol*. 2022; 60(1): e0160721. doi: 10.1128/JCM.01607-21
 204. Hoenigl M, Sprute R, Egger M, et al. The Antifungal Pipeline: Fosmanogepix, Ibrexafungerp, Olorofim, Opelconazole, and Rezafungin. *Drugs*. 2021; 81(15): 1703-1729. doi: 10.1007/s40265-021-01611-0
 205. Dos Santos Ramos MA, Da Silva PB, Spósito L, et al. Nanotechnology-based drug delivery systems for control of microbial biofilms: a review. *Int J Nanomedicine*. 2018; 13: 1179-1213. doi: 10.2147/IJN.S146195
 206. Fernández de Ullivarri M, Arbulu S, Garcia-Gutierrez E, Cotter PD. Antifungal Peptides as Therapeutic Agents. *Front Cell Infect Microbiol*. 2020; 10: 105. doi: 10.3389/fcimb.2020.00105
 207. Basavegowda N, Baek KH. Combination Strategies of Different Antimicrobials: An Efficient and Alternative Tool for Pathogen Inactivation. *Biomedicines*. 2022; 10(9). doi: 10.3390/biomedicines10092219
 208. Fleitas Martínez O, Cardoso MH, Ribeiro SM, Franco OL. Recent Advances in Anti-virulence Therapeutic Strategies With a Focus on Dismantling Bacterial Membrane Microdomains, Toxin Neutralization, Quorum-Sensing Interference and Biofilm Inhibition. *Front Cell Infect Microbiol*. 2019; 9: 74. doi: 10.3389/fcimb.2019.00074
 209. Burks C, Darby A, Gómez Londoño L, Momany M, Brewer MT. Azole-resistant *Aspergillus fumigatus* in the environment: Identifying key reservoirs and hotspots of antifungal resistance. *PLoS Pathog*. 2021; 17(7): e1009711. doi: 10.1371/journal.ppat.1009711
 210. David S, Caballero JD, Couto N, et al. Monitoring antimicrobial resistance trends from global genomics data: amr.watch. *PLOS Glob public Heal*. 2025; 5(11): e0005256. doi: 10.1371/journal.pgph.0005256
 211. Sithole MN, Khan MR, Mohammed HA, Khan RA, Naik K, Choonara YE. A Systematic Review on Vaccine Developmental Approaches: Evaluating Efficacy, and Addressing Challenges of Infectious Diseases in the Post-COVID-19 Era. *Virus Res*. Published online 2026: 199720. doi: <https://doi.org/10.1016/j.virusres.2026.199720>
 212. Almatroudi A. Biofilm Resilience: Molecular Mechanisms Driving Antibiotic Resistance in Clinical Contexts. *Biology (Basel)*. 2025; 14(2). doi: 10.3390/biology14020165
 213. Tanwar S, Kalra S, Bari VK. Insights into the role of sterol metabolism in antifungal drug resistance: a mini-review. *Front Microbiol*. 2024; 15: 1409085. doi: 10.3389/fmicb.2024.1409085
 214. Alves V, Zamith-Miranda D, Frases S, Nosanchuk JD. Fungal Metabolomics: A Comprehensive Approach to Understanding Pathogenesis in Humans and Identifying Potential Therapeutics. *J Fungi*. 2025; 11(2). doi: 10.3390/jof11020093
 215. Whitehead AJ, Woodring T, Klein BS. Immunity to fungi and vaccine considerations. *Cell Host Microbe*. 2024; 32(10): 1681-1690. doi: 10.1016/j.chom.2024.09.011
 216. Hou Y, Chen M, Bian Y, Zheng X, Tong R, Sun X. Advanced subunit vaccine delivery technologies: From vaccine cascade obstacles to design strategies. *Acta Pharm Sin B*. 2023; 13(8): 3321-3338. doi: 10.1016/j.apsb.2023.01.006
 217. Huang X, Moses M, Nie L, Wandera EA, Chen C. Diagnostics for human pathogenic fungal infections: Current status and future prospects. *hLife*. 2026; 4(3): 135-164. doi: <https://doi.org/10.1016/j.hlife.2025.11.005>
 218. Williams CC, Gregory JB, Usher J. Understanding the clinical and environmental drivers of antifungal resistance in the One Health context. *Microbiology*. 2024; 170(10). doi: 10.1099/mic.0.001512
 219. Horvat O, Kovačević Z. Human and Veterinary Medicine Collaboration: Synergistic Approach to Address Antimicrobial Resistance Through the Lens of Planetary Health. *Antibiot (Basel, Switzerland)*. 2025; 14(1). doi: 10.3390/antibiotics14010038